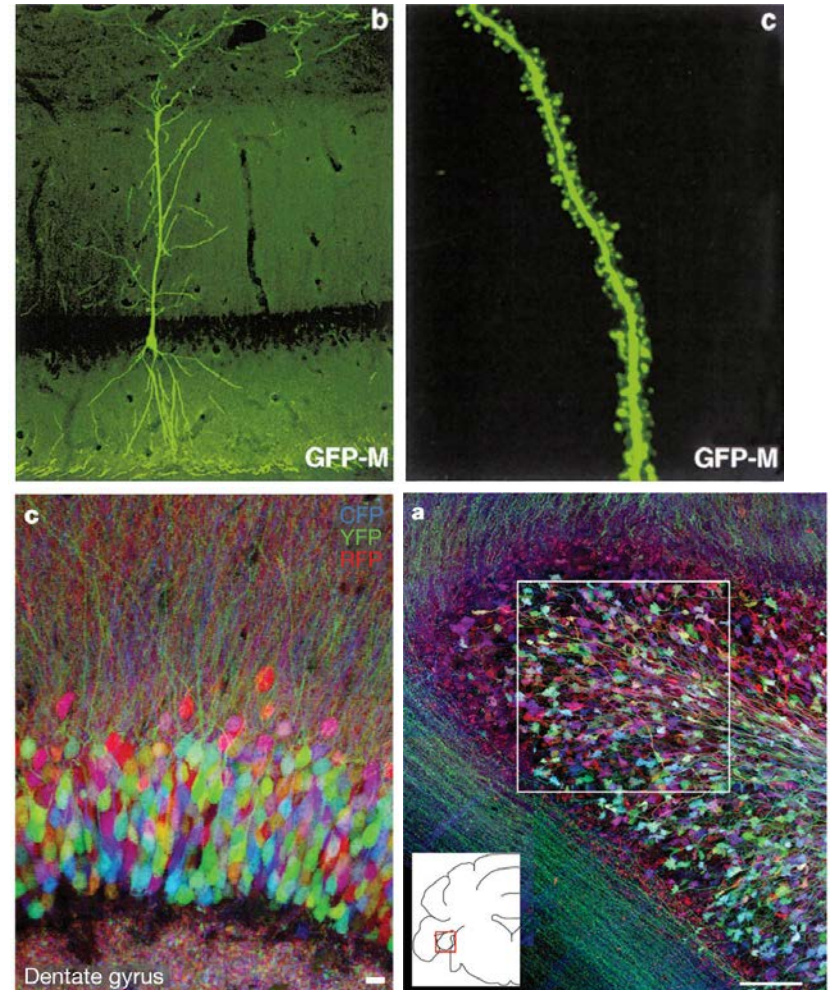
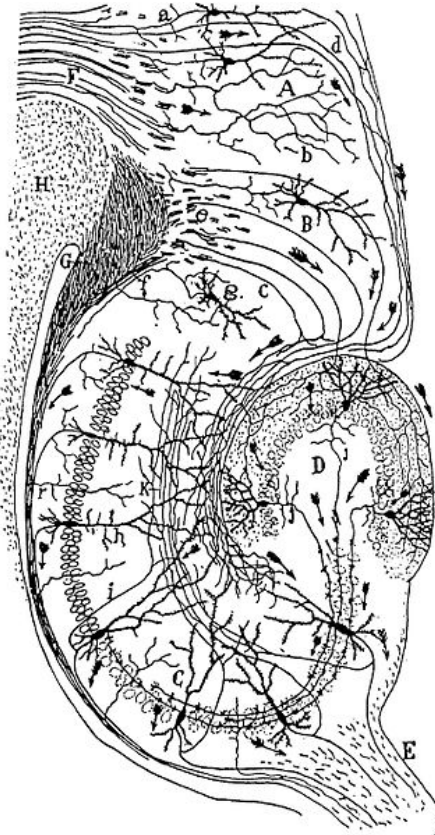
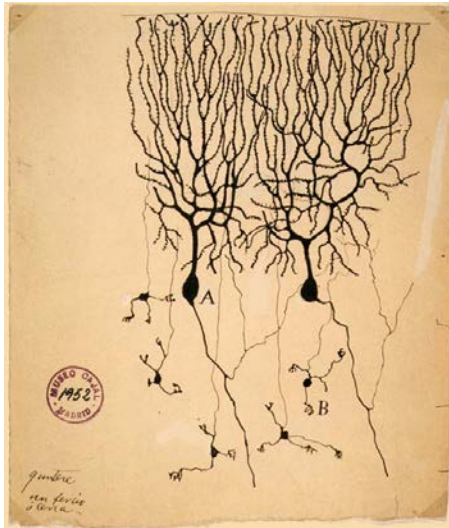


Introduction to Genetic Tools in Neurobiology & NS201B Overview



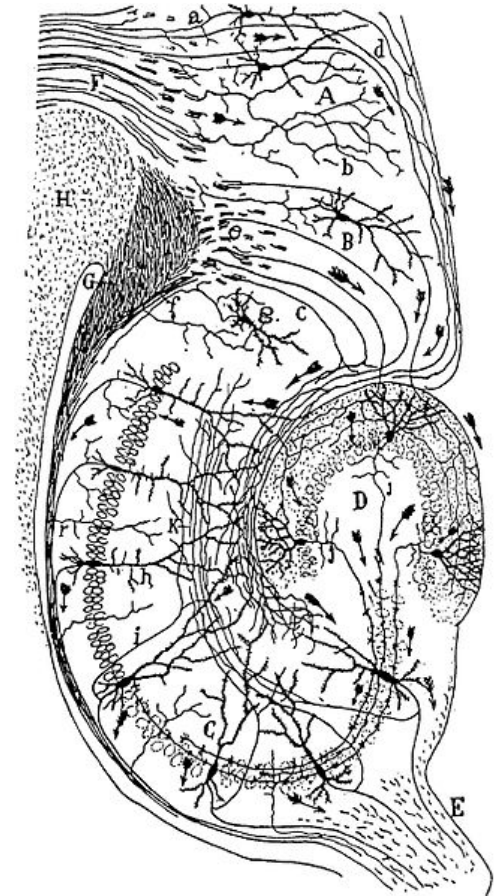
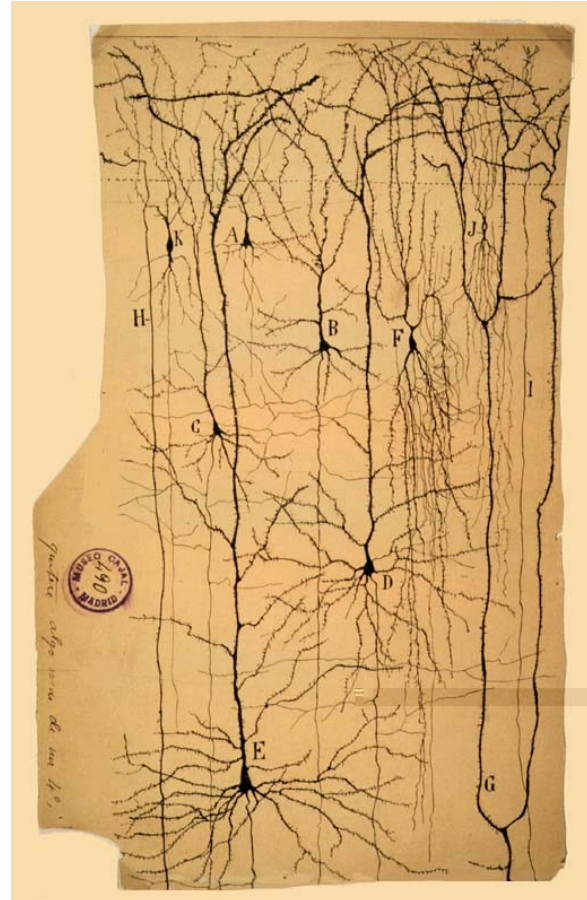
Eric J. Huang, Professor of Pathology
Eric.huang2@ucsf.edu

The Neuron Doctrine

The fundamental unit in the nervous system



Santiago Ramón y Cajal 1852-1934



1906 Nobel Prize in Physiology or Medicine (shared with Camillo Golgi)

The Polarity Hypothesis

Directional flows in dendrites, soma and axons

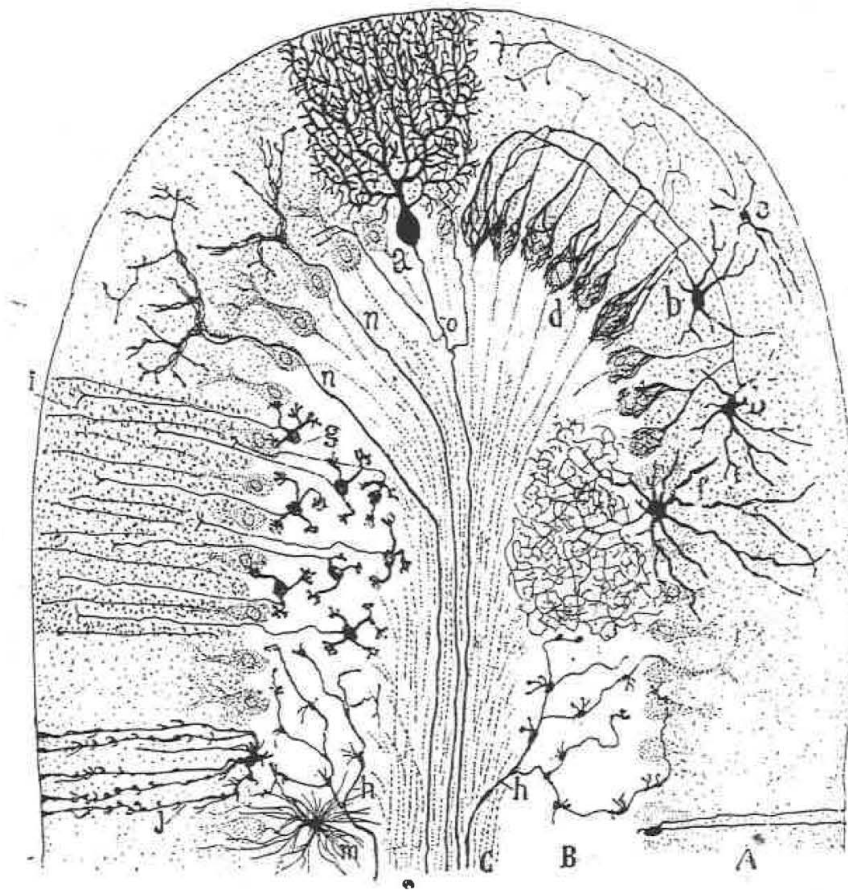


FIG. 28. SEMIDIAGRAMMATIC TRANSVERSE SECTION OF A CEREBELLAR CONVOLUTION OF A MAMMAL. *A*, molecular layer; *B*, granular layer; *C*, layer of white matter; *a*, Purkinje cell with its dendrites spread out in the plane of section; *b*, small stellate cells of the molecular layer; *d*, descending terminal arborizations embracing the cells of Purkinje; *e*, superficial stellate cell; *f*, large stellate cell of the granule layer; *g*, granules with their ascending axons bifurcating at *i*; *h*, mossy fibres; *j*, tufted neuroglia cell; *n*, climbing fibres; *m*, neuroglia cell of the granule layer.

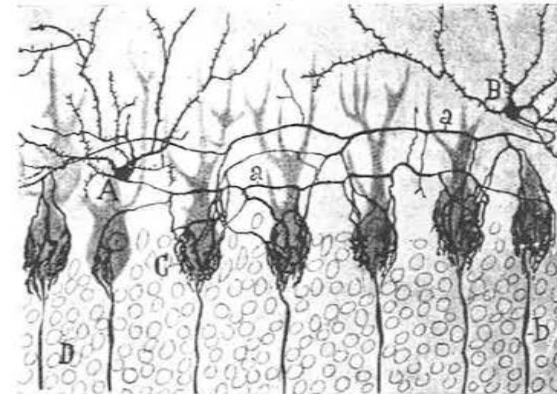


FIG. 26. TRANSVERSE SECTION OF A CEREBELLAR LAMELLA. Semidiagrammatic. *A* and *B*, stellate cells of the molecular layer (basket cells), of which the axon (*a*) produces terminal nests about the cells of Purkinje (*C*): *b*, axon of the Purkinje cell.

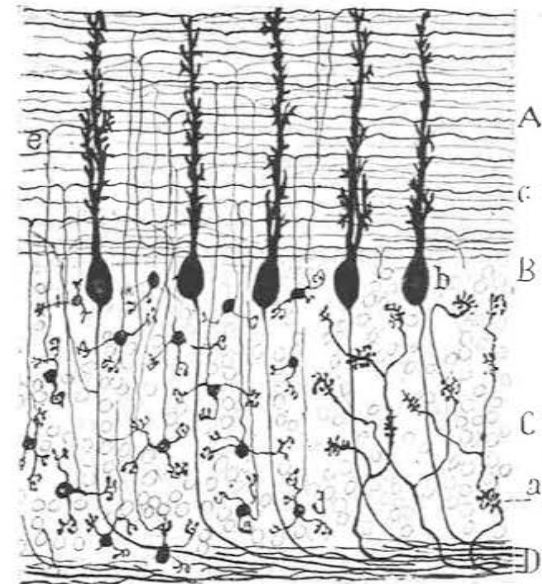


FIG. 27. LONGITUDINAL SECTION OF A CEREBELLAR CONVOLUTION. *A*, molecular layer; *B*, layer of Purkinje cells; *C*, granular layer; *D*, white matter; *a*, tuft of a mossy fibre; *b*, body of a Purkinje cell; *c*, parallel fibres; *d*, granule cell with its ascending axon; *e*, division of this axon. (Semidiagrammatic.)

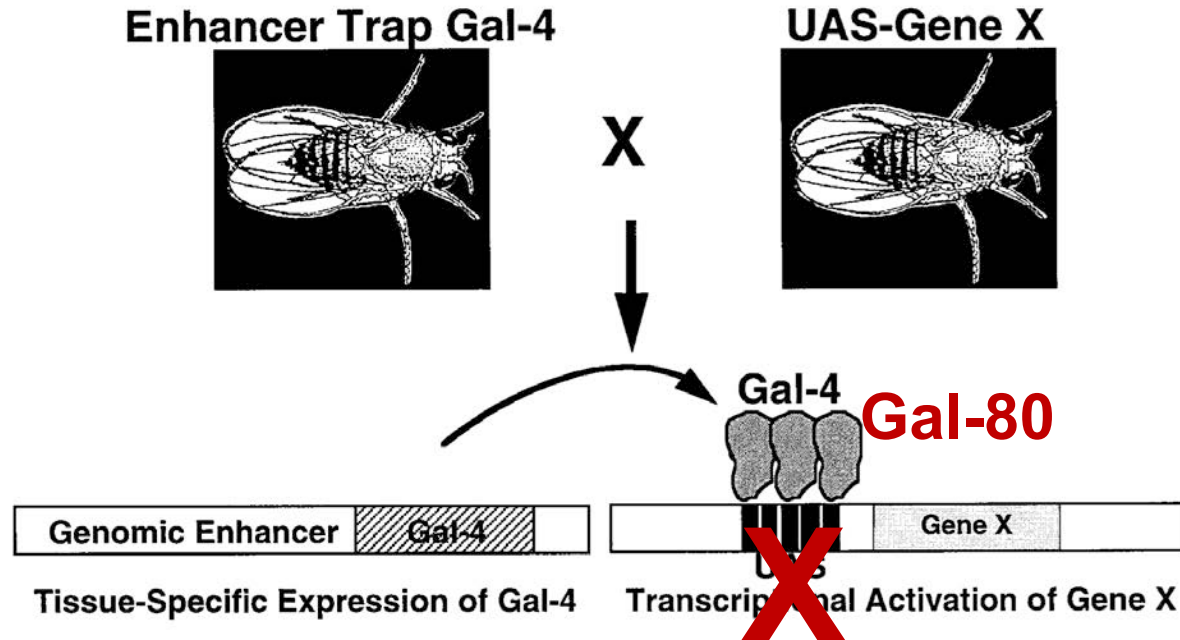
Introduction to Genetic Tools in Neurobiology

- **Genetic approaches to investigate the complexity of the nervous system**
 - Lineage tracing of neuronal origin
 - Cell biology of neuron
 - Neural circuits and connectivity
- **Model organisms**
 - *Caenorhabditis elegans*
 - *Drosophila melanogaster*
 - *Mus musculus*
 - *Rattus rattus*
- **Technical approaches**
 - Transgenic
 - Conditional gene targeting
- **Functional outcomes**
 - Loss of function vs. Gain of function
 - Cell type-specific vs. circuit-dependence

Take Home Message:
Genetic tools allow temporal and spatial controls to study gene and circuit functions in neurobiology

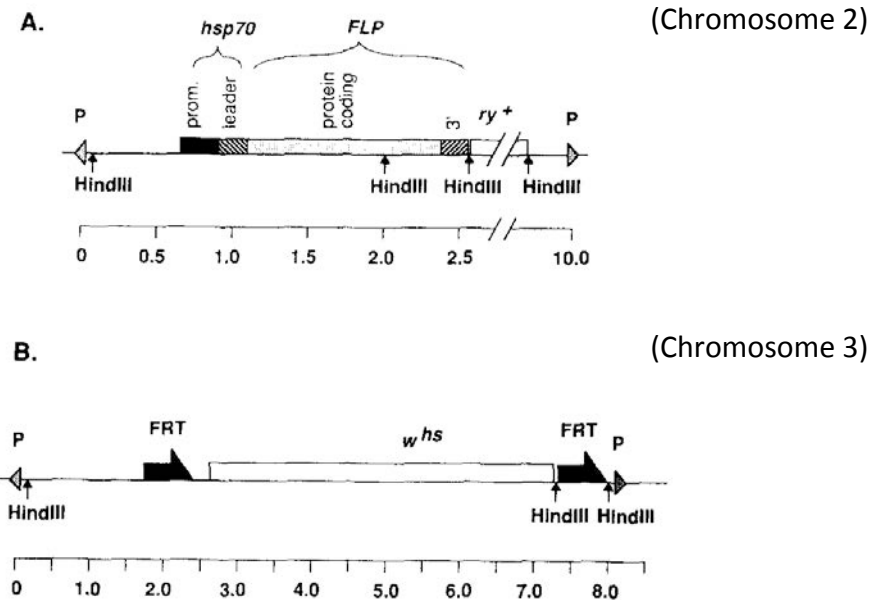
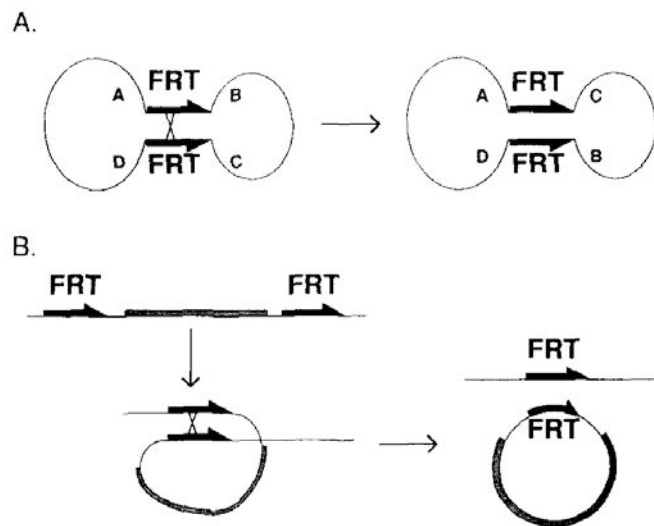
Genetic tools in neurobiology research in *Drosophila*

- The “GAL4-UAS” technique – misexpression of gene
- The “FLP/FRT” technique – germline mosaics
- The MARCM system – single cells & clonal analyses



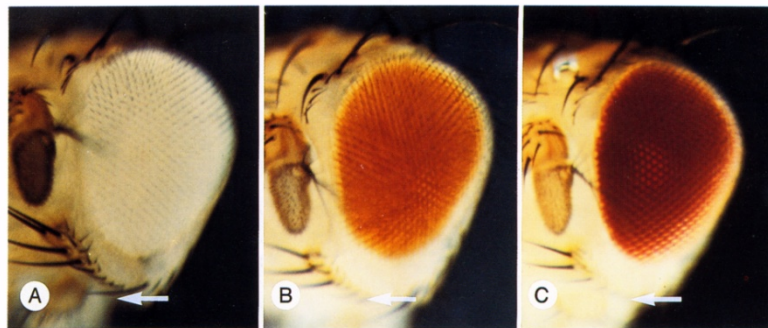
The “FLP/FRT” technique

- FLP: a recombinase encoded by the yeast 2 μ m plasmid
- FRT: FLP recombination target present in the 2 μ m plasmid as 599bp inverted repeats



(Golic and Linquist, Cell 1989)

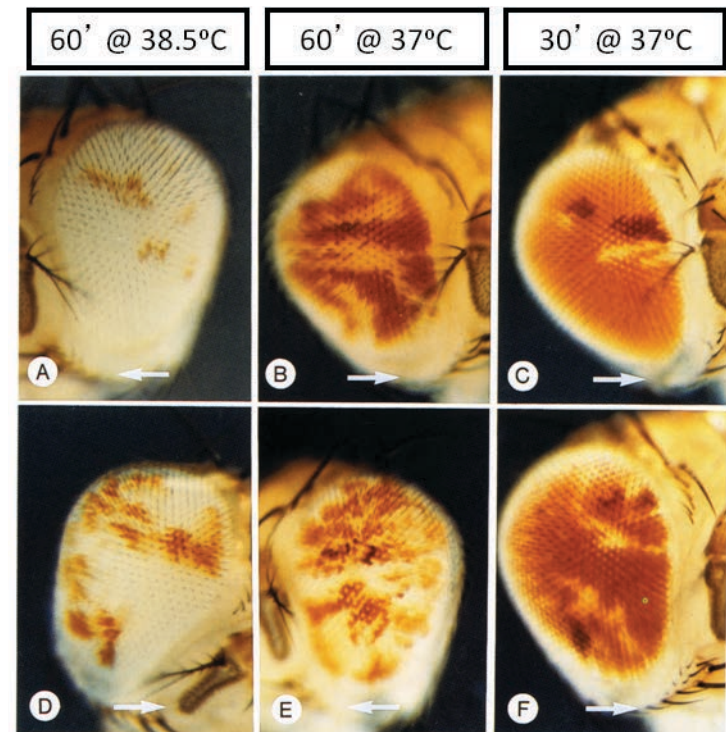
The “FLP/FRT” technique



w/w
loss of eye
color;
'white'
eye

w/hsp70-W+
One copy
of eye
color
transgene

hsp70-W+/hsp70-W+
two copies of
eye
color
transgene



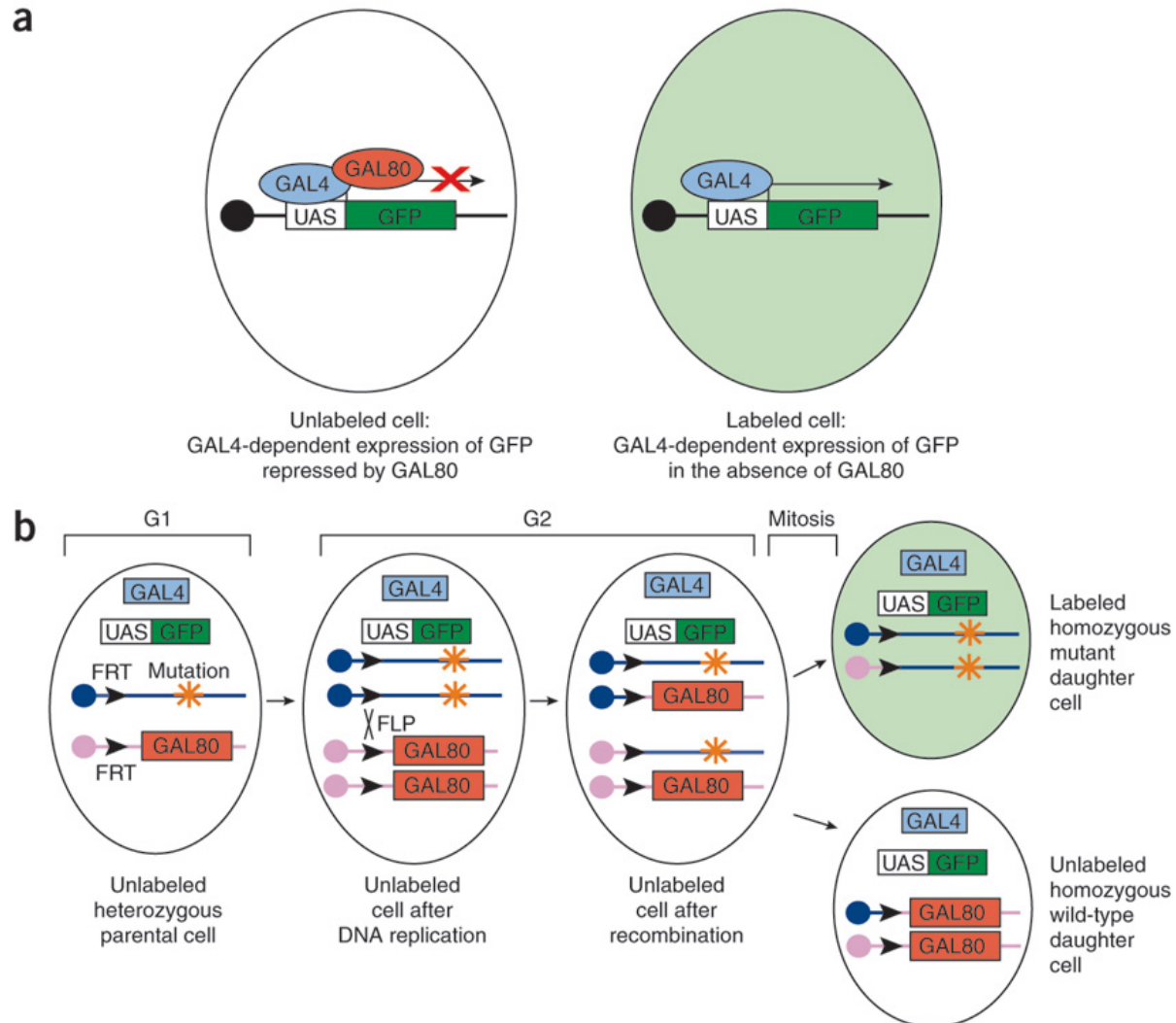
(Golic and Linquist, Cell 1989)

Mosaic Analysis with a Repressible Cell Marker (MARCM)

- **Positively marks a small population of wild type or mutant cells**
- **Generate homozygous mutant cells from heterozygous precursors via mitotic recombination**
- **MARCM-ready flies: GAL4-UAS, GAL80 and FLP/FRT**
- **Applications:**
 - Lineage analysis
 - Investigating gene function in single or small populations of cells
 - Neuronal circuit tracing
 - Growth cone signaling
 - Axon pruning

(Lee T, Luo L, Neuron, 1999)

Mosaic Analysis with a Repressible Cell Marker (MARCM)

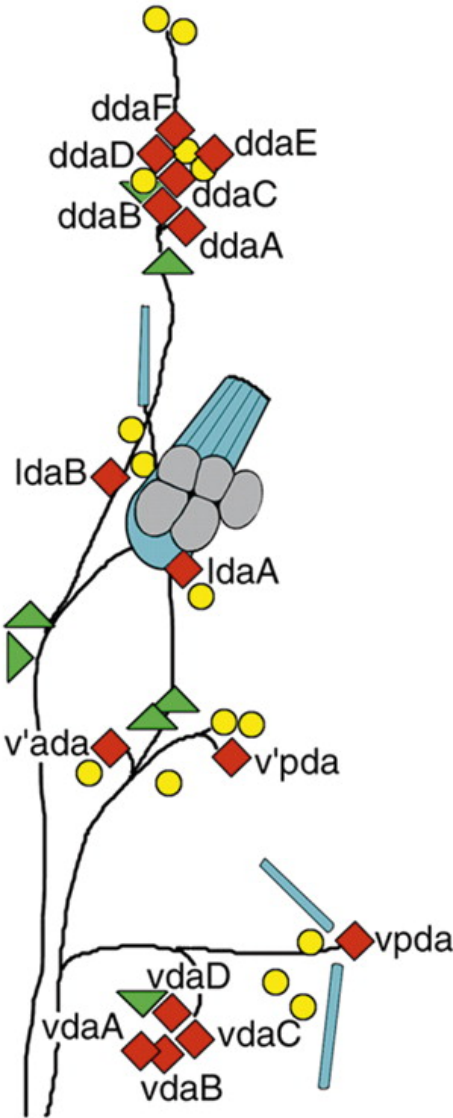


Example: MARCM used to analyze morphology

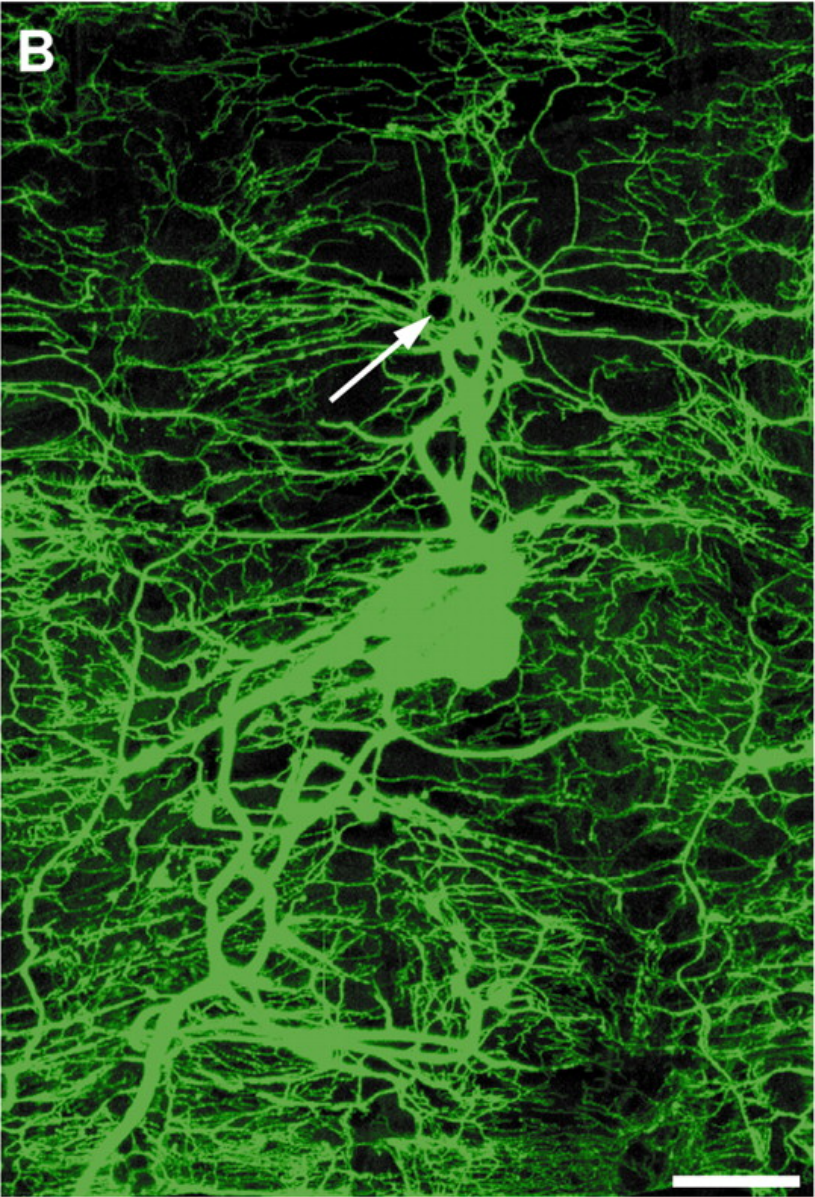
- Assessment of cell autonomous gene function
 - Single cells labeled
 - Early lethal genes can be analyzed in specific cell types at later stages of development
- Cut (ct) gene
 - Transcription factor (homeobox gene)
 - Mutants are embryonic lethal
 - Expressed in Drosophila Peripheral Nervous System (PNS) sensory neurons

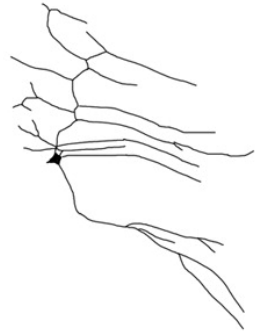
Drosophila da neuron clusters
(from one segment)

A

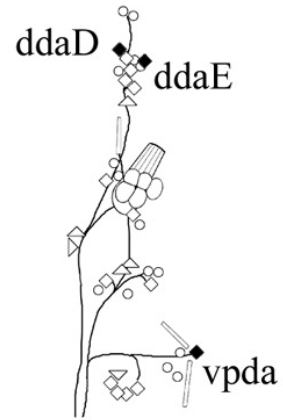
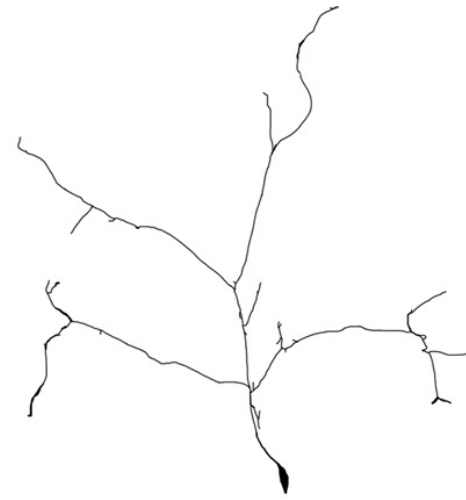


*Gal4*¹⁰⁹⁽²⁾⁸⁰, UAS-mCD8-GFP (all da neurons)

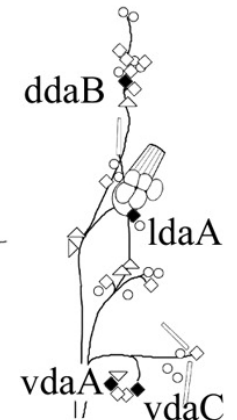
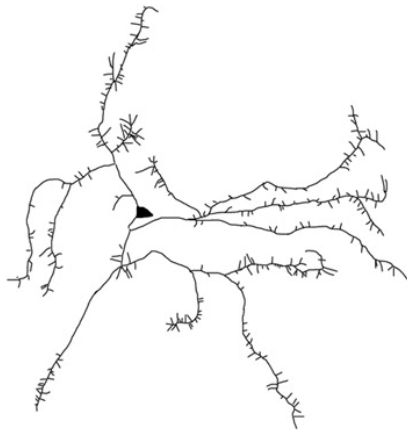


C

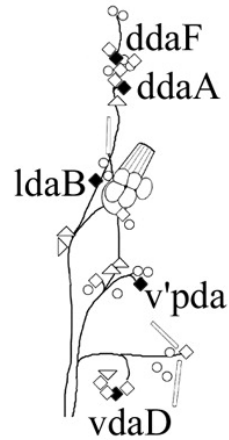
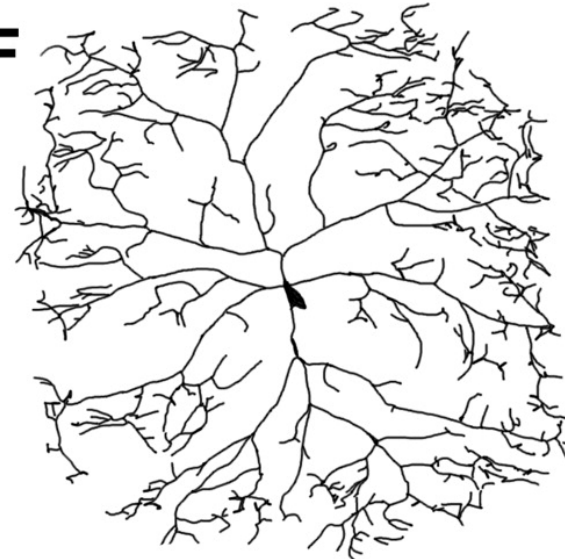
ddaE (class I)

**D**

ddaB (class II)

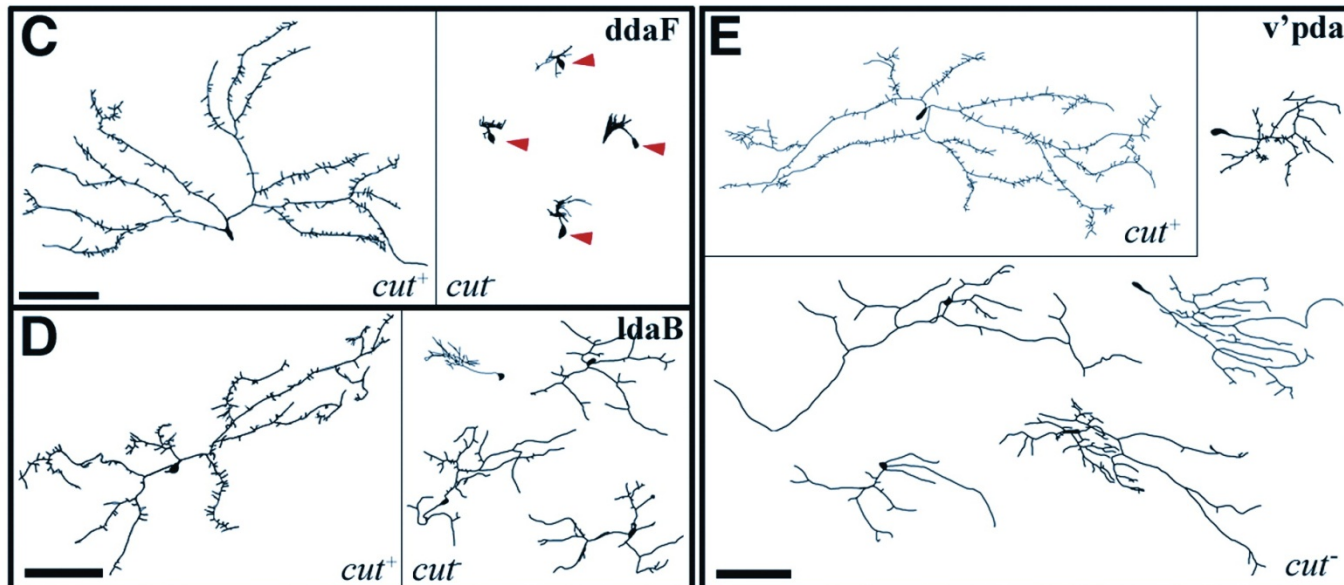
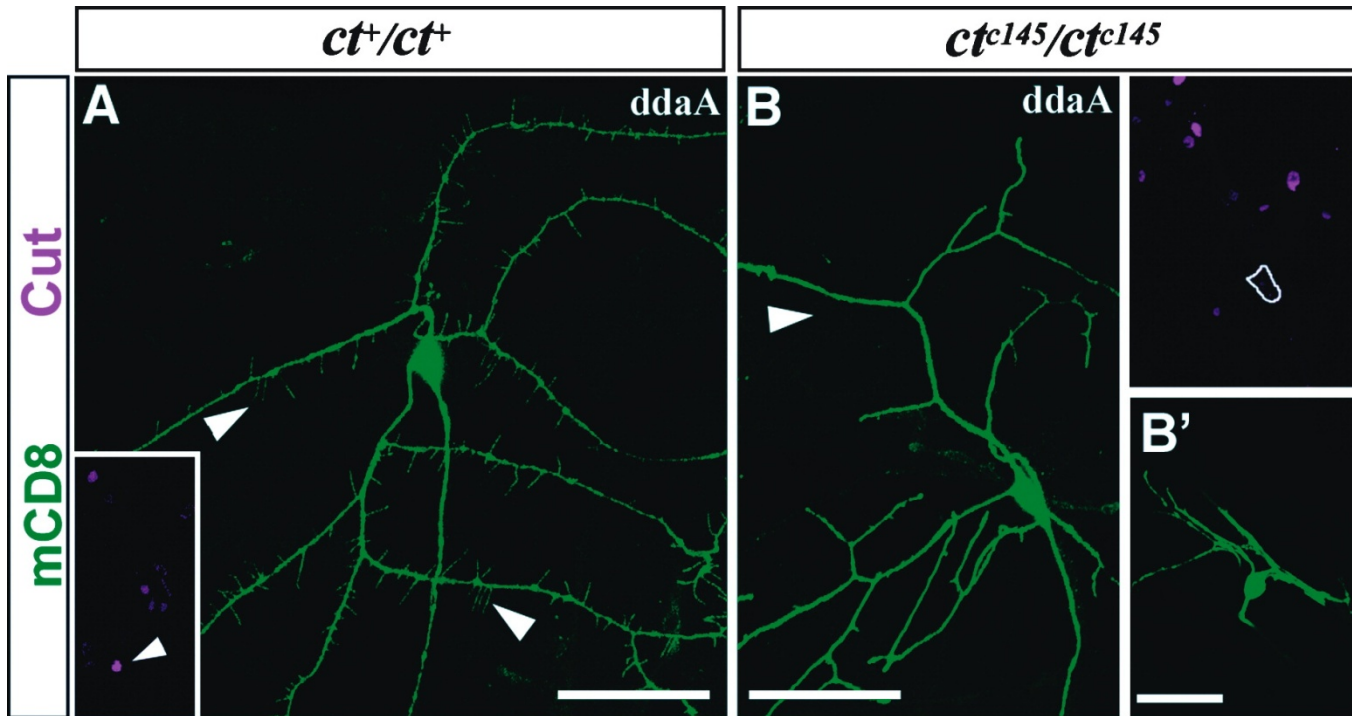
**E**

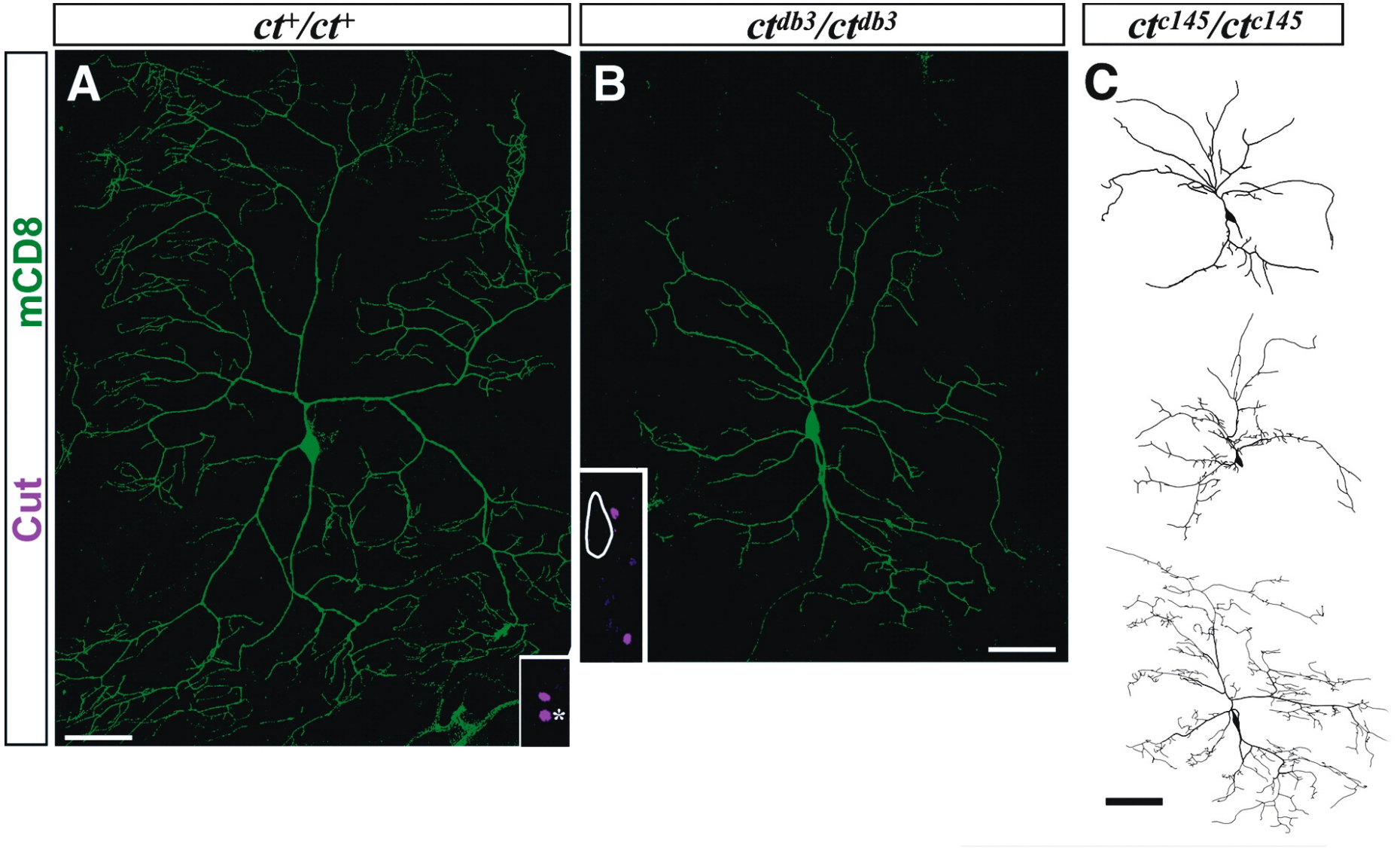
ddaA (class III)

**F**

ddaC (class IV)





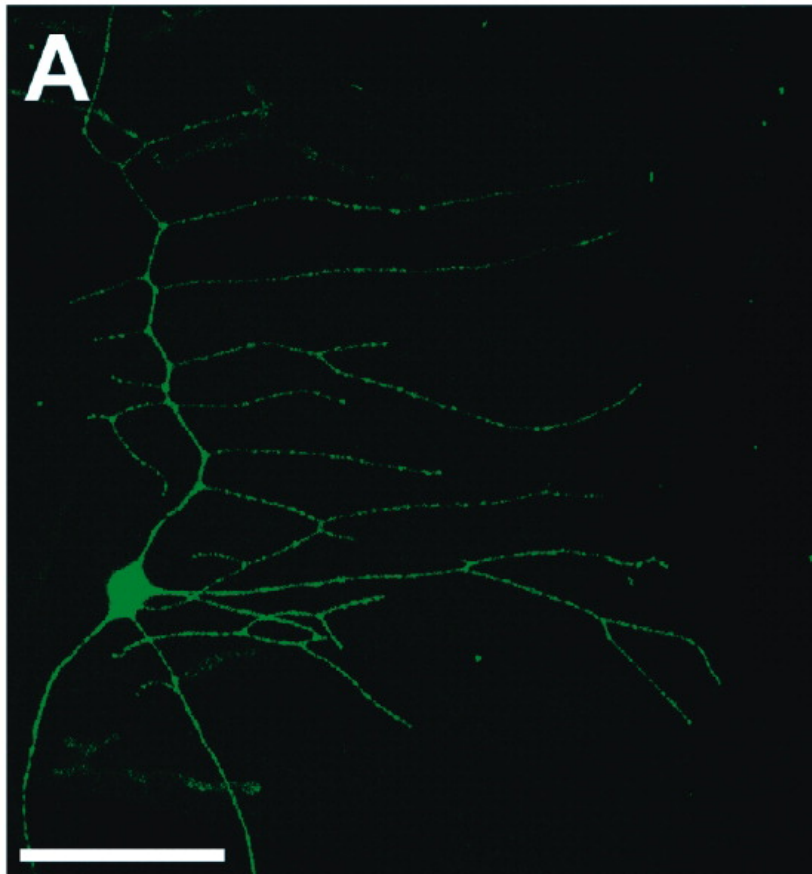


ct⁺/ct⁺

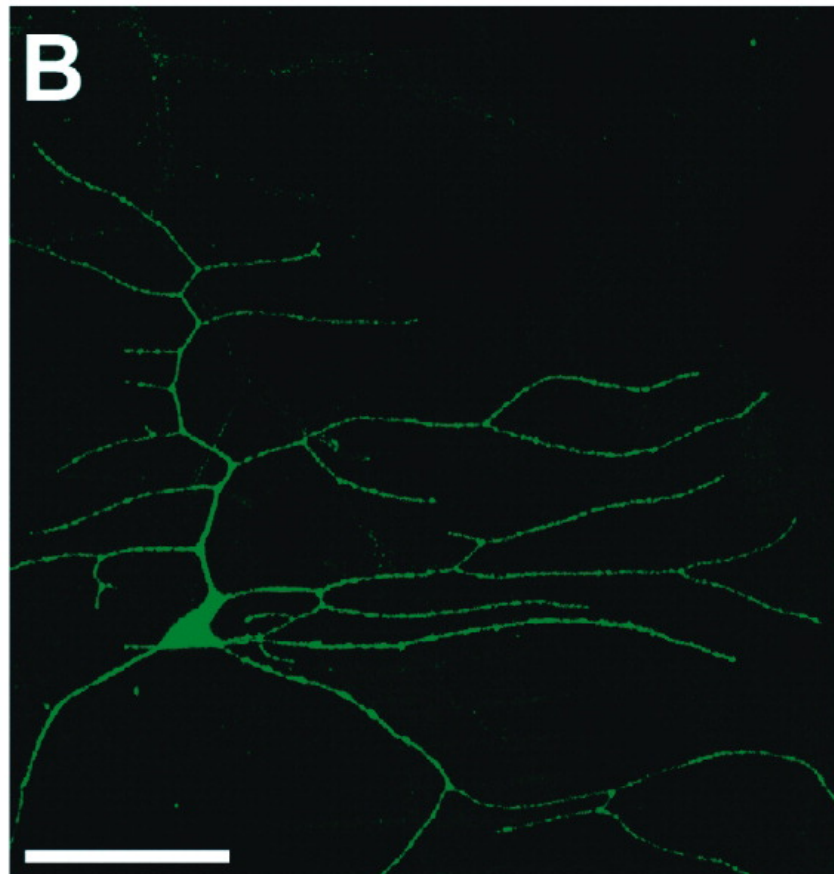
ct^{c145}/ct^{c145}

mCD8

A

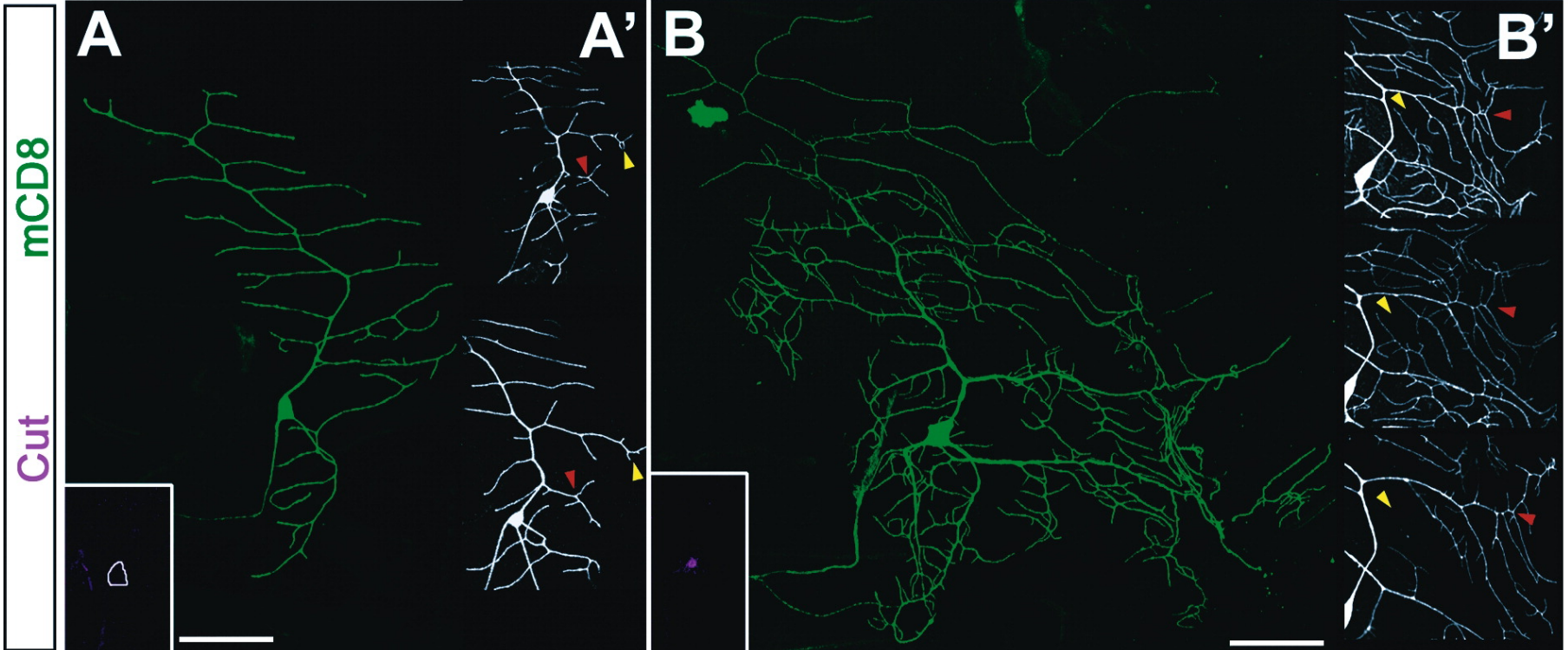


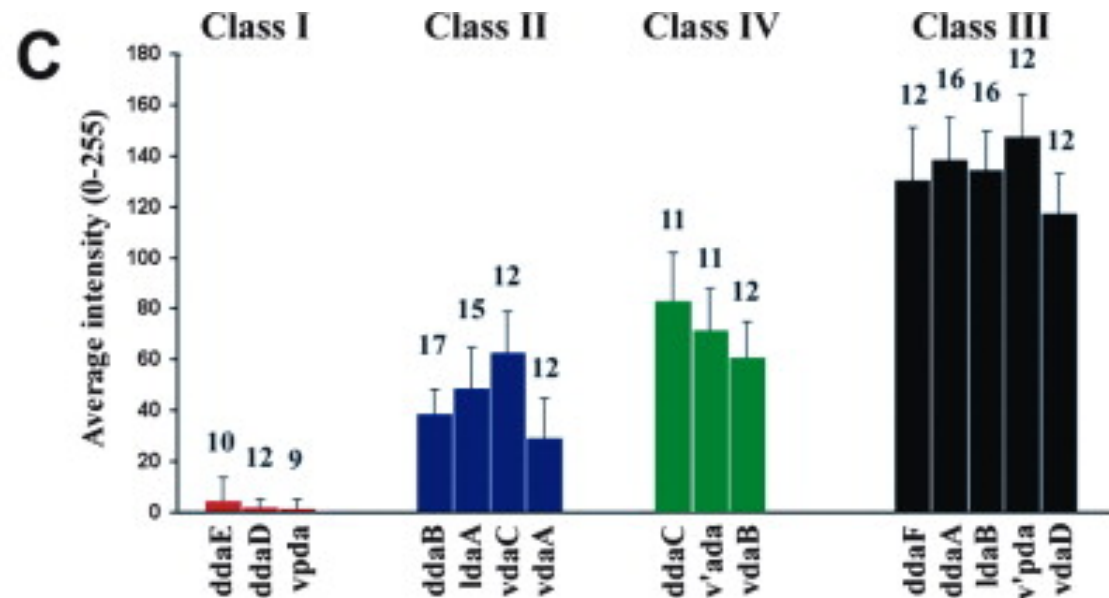
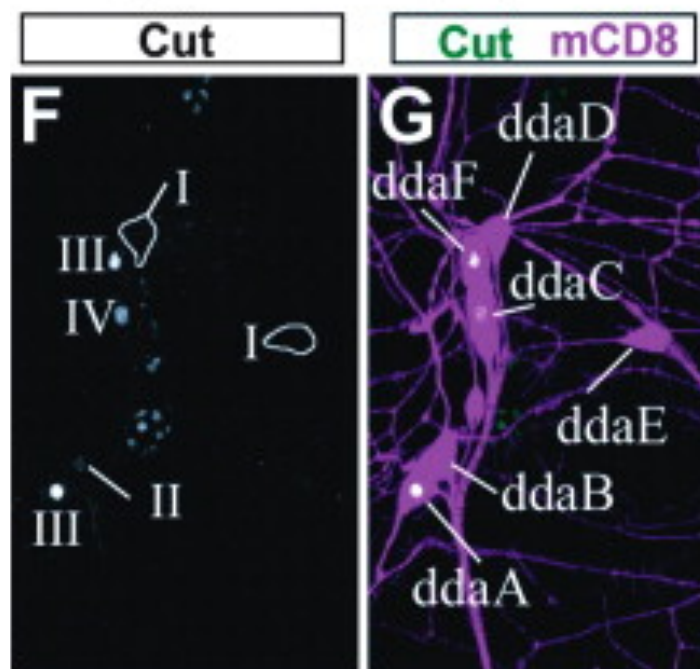
B



Gal4²²¹,UASmCD8GFP x w¹¹¹⁸

Gal4²²¹,UASmCD8GFP x UAS-cut





H

| | Class I | Class II | Class IV | Class III |
|----------------------|--------------|----------|-----------------------|------------------|
| Cut Immunoreactivity | Undetectable | Low | Medium | High |
| Dendrite morphology | Simple | Simple | Complex space-filling | Complex "spiked" |

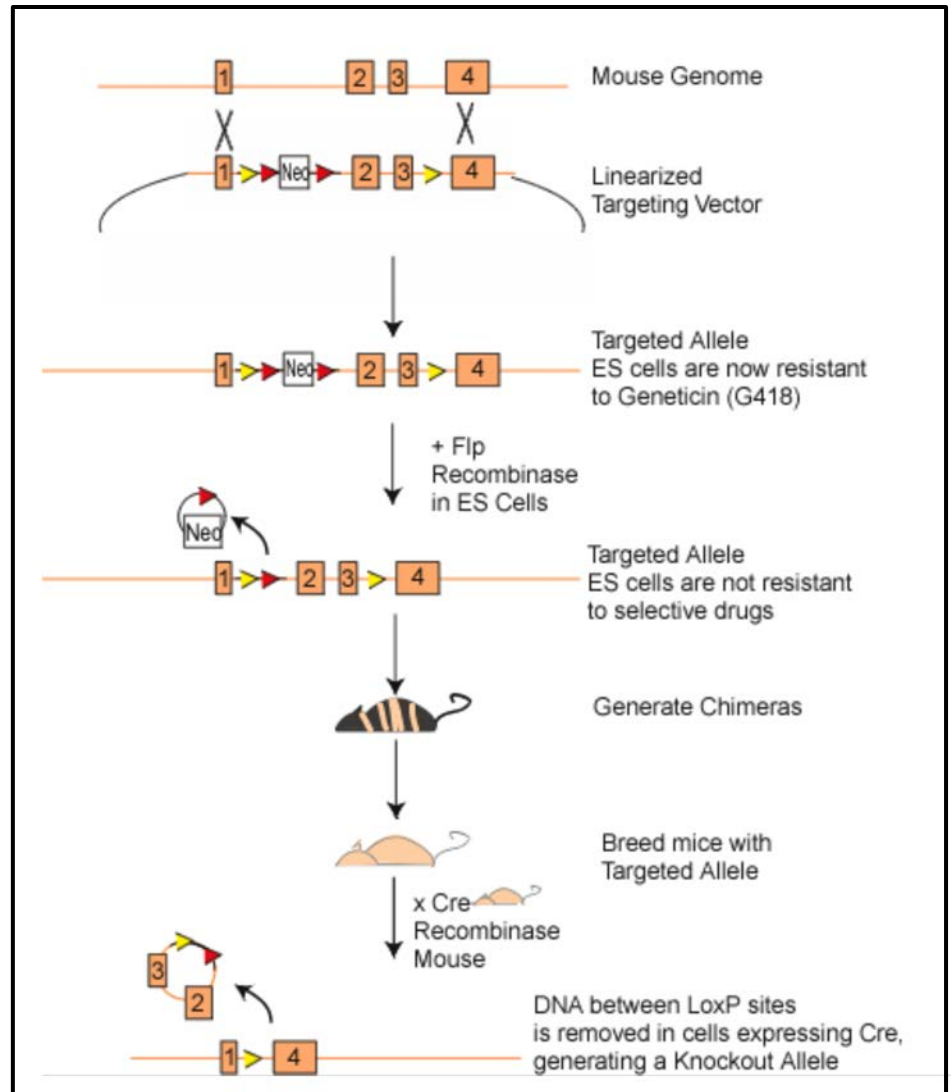
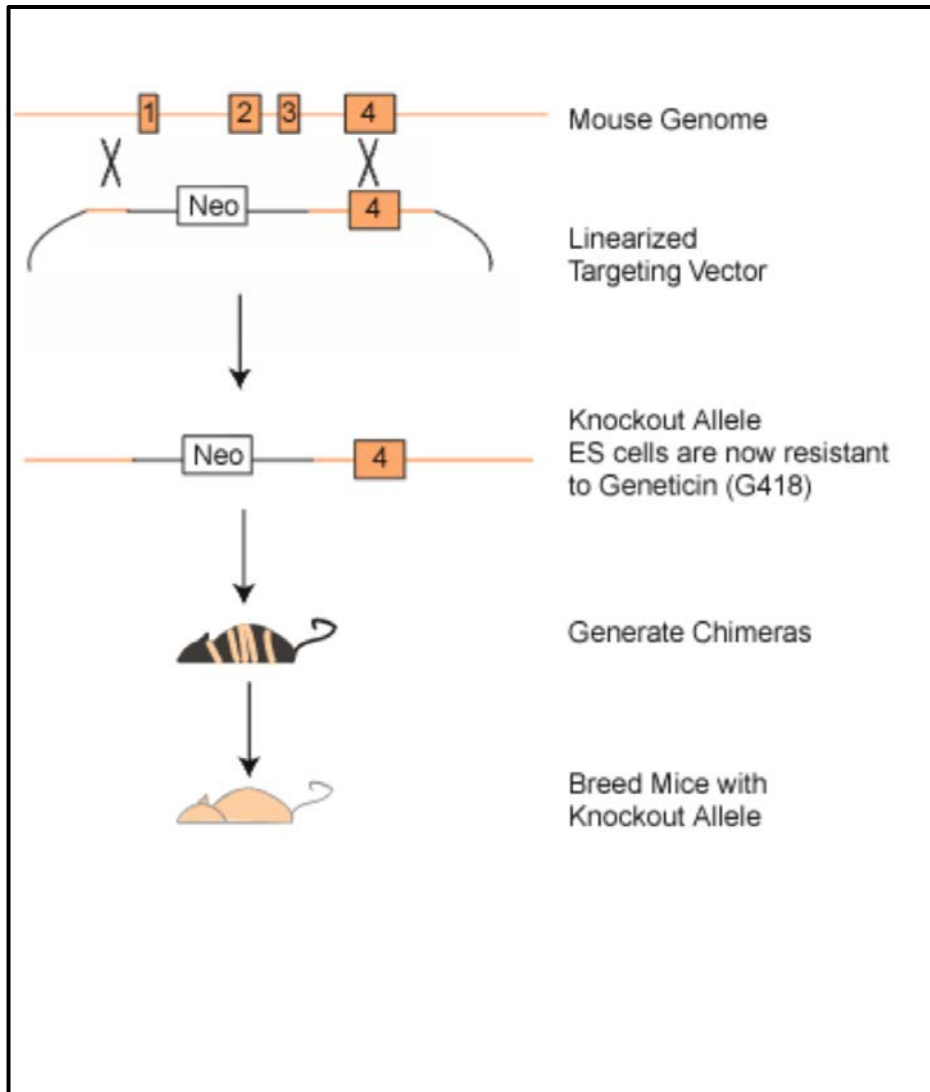
Limitations of MARCM

- **Timing of GAL80 elimination:** due to protein perdurance, MARCM can only be used to reliably label single cells 24-48 hours after the induction of mitotic recombination
- **Maternal GAL80 contribution:** limits the efficacy of the MARCM system in studying early embryo development
- **Gene of interest:** the high abundance and perdurance of protein may interfere with interpretations
- **Some mutant cells may not be labeled by Gal4-UAS:** Gal4 is only expressed in subset of cells, but hs-FLP/FRT recombination is ubiquitous

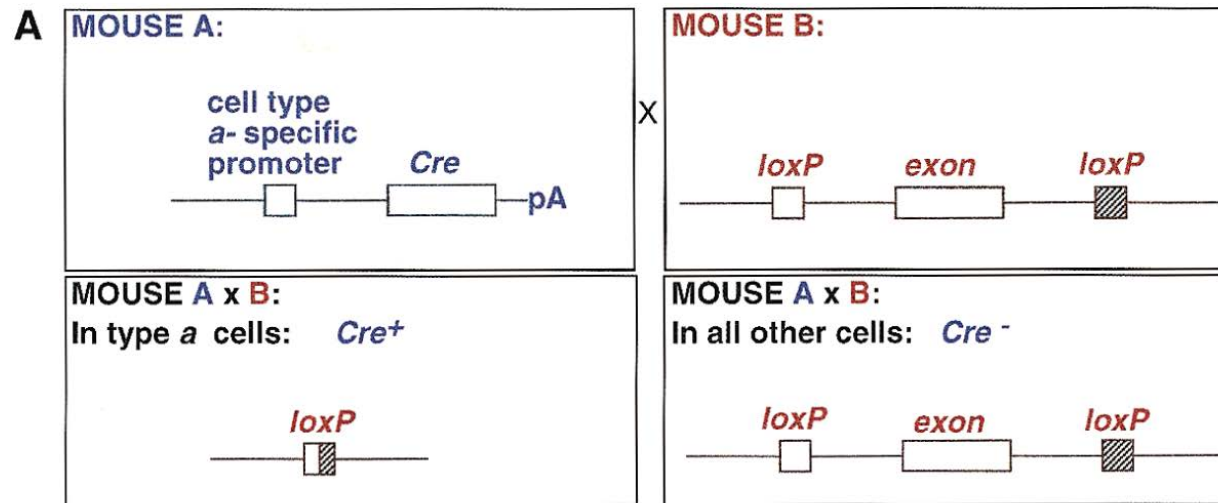
Genetic tools for neuronal functions and neural circuits in mouse brain

- **Transgenic targeting of specific neuron type**
 - *cis*-regulatory elements as drivers
 - Bacterial artificial chromosome (BAC) transgenics
 - Gene targeting (“knock-in”) w/ or w/o internal ribosomal entry site (IRES)
 - Enhancer trap – random insertion of target genes in the genome under the control of a minimal promoter
- **Binary expression strategy – Cre-loxP, FLP-FRT & Tet-on/Tet-off systems**
 - Cre drivers
 - Cre reporter lines
- **GFP reporter and its variants**

Schematic diagrams of conventional and conditional knockout in mice

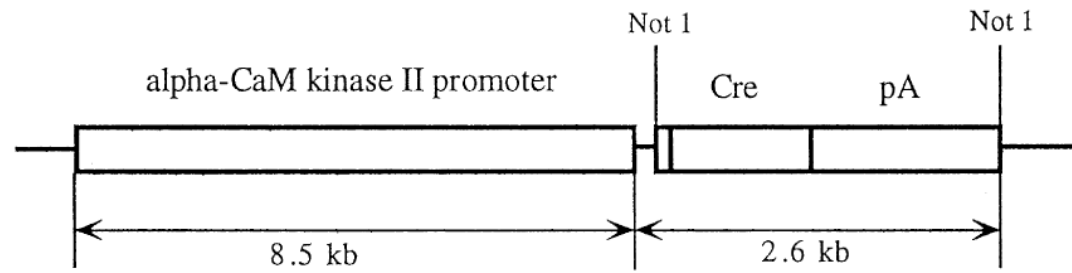


Strategy for cell type-specific gene knockout

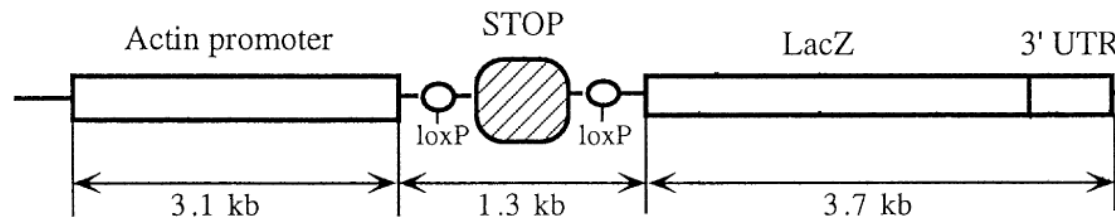


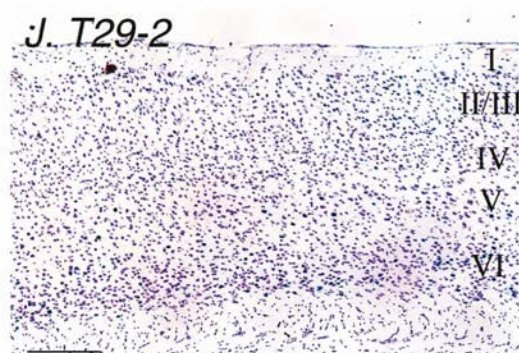
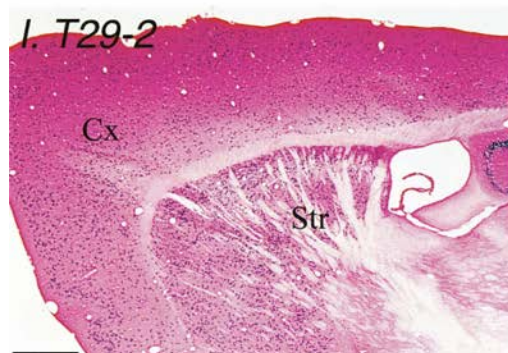
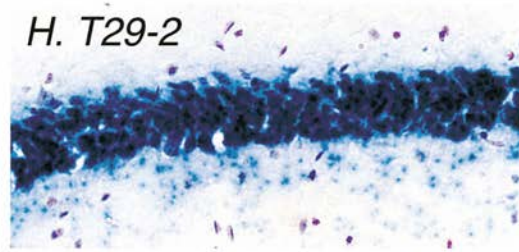
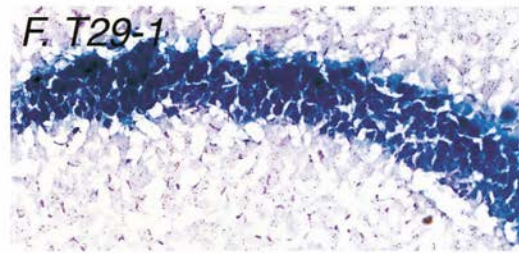
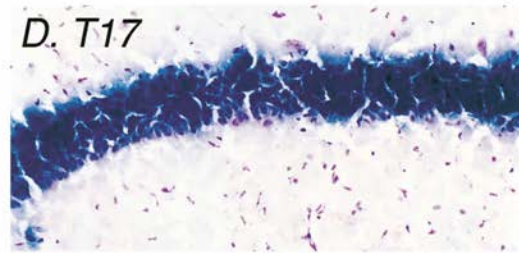
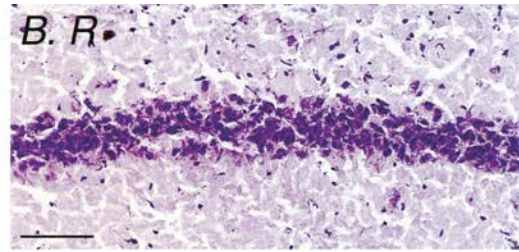
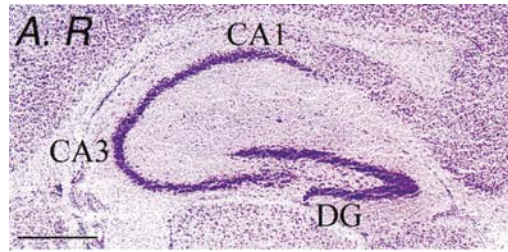
Strategy for cell type-specific gene knockout

A. Construct (pJT-CRE) for production of *Cre* Mouse



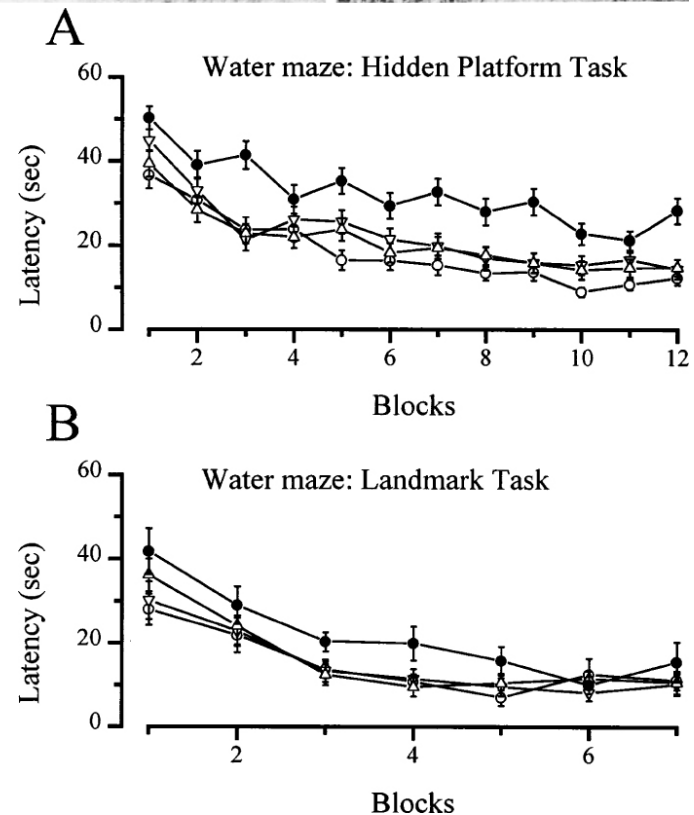
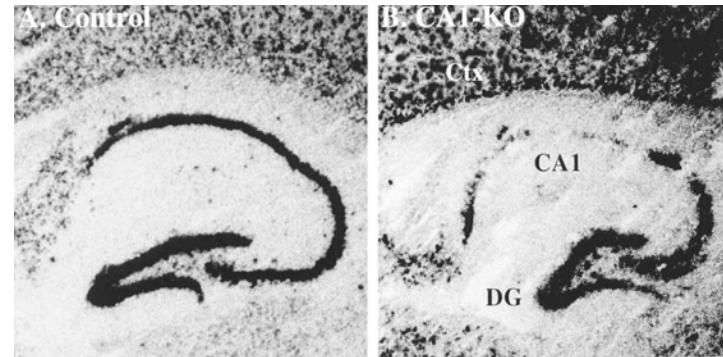
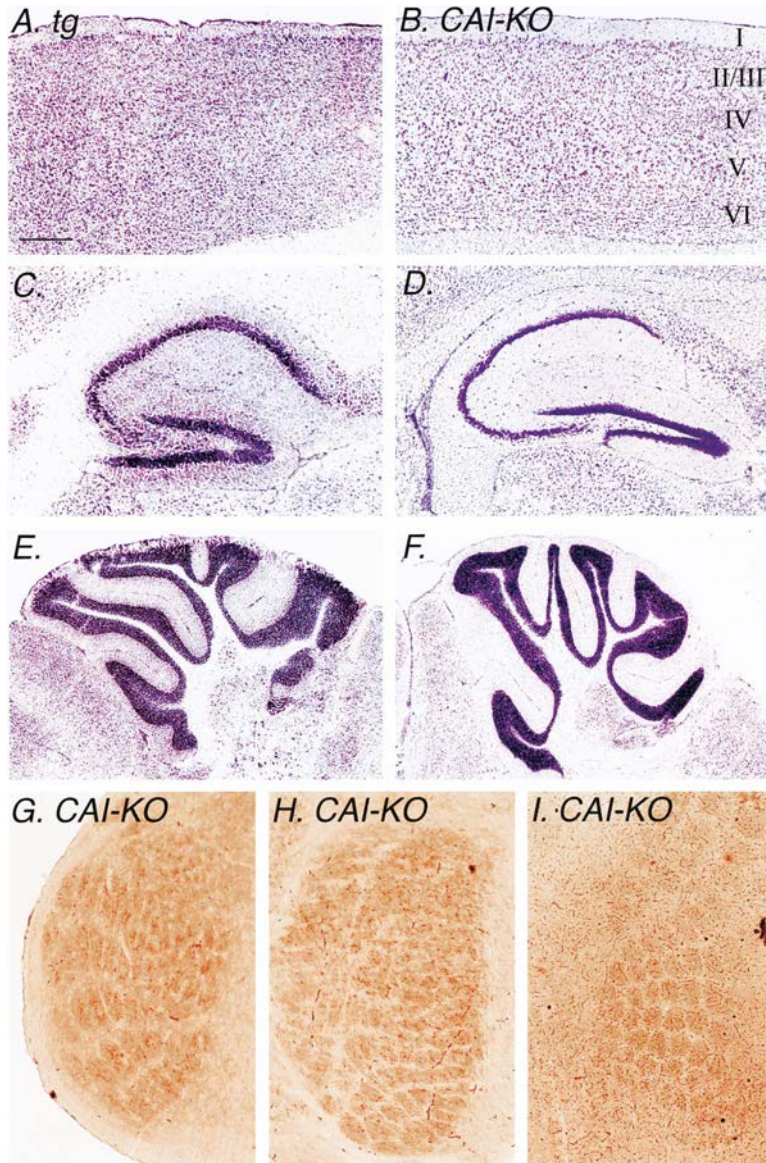
B. Construct (pcAct-XstopX-LacZ) for production of *Reporter* Mouse





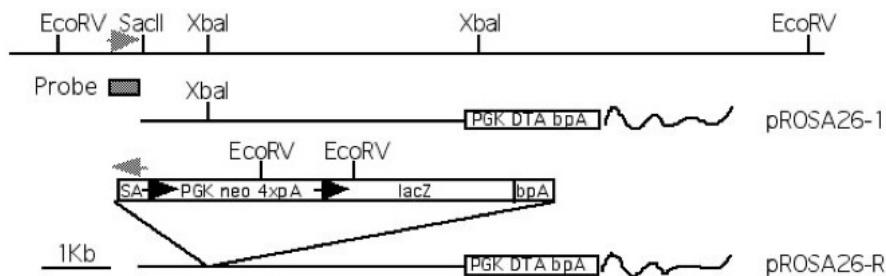
(Tsien et al., Cell 1996a)

Activity dependent modification of CA1 synapse, mediated by NMDA receptor, is required for spatial learning

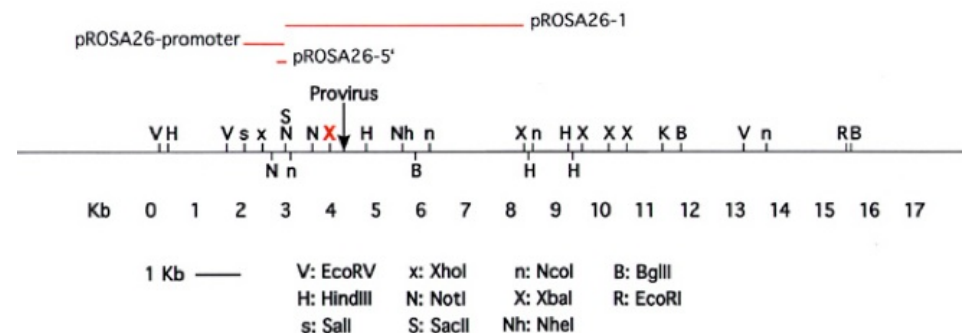


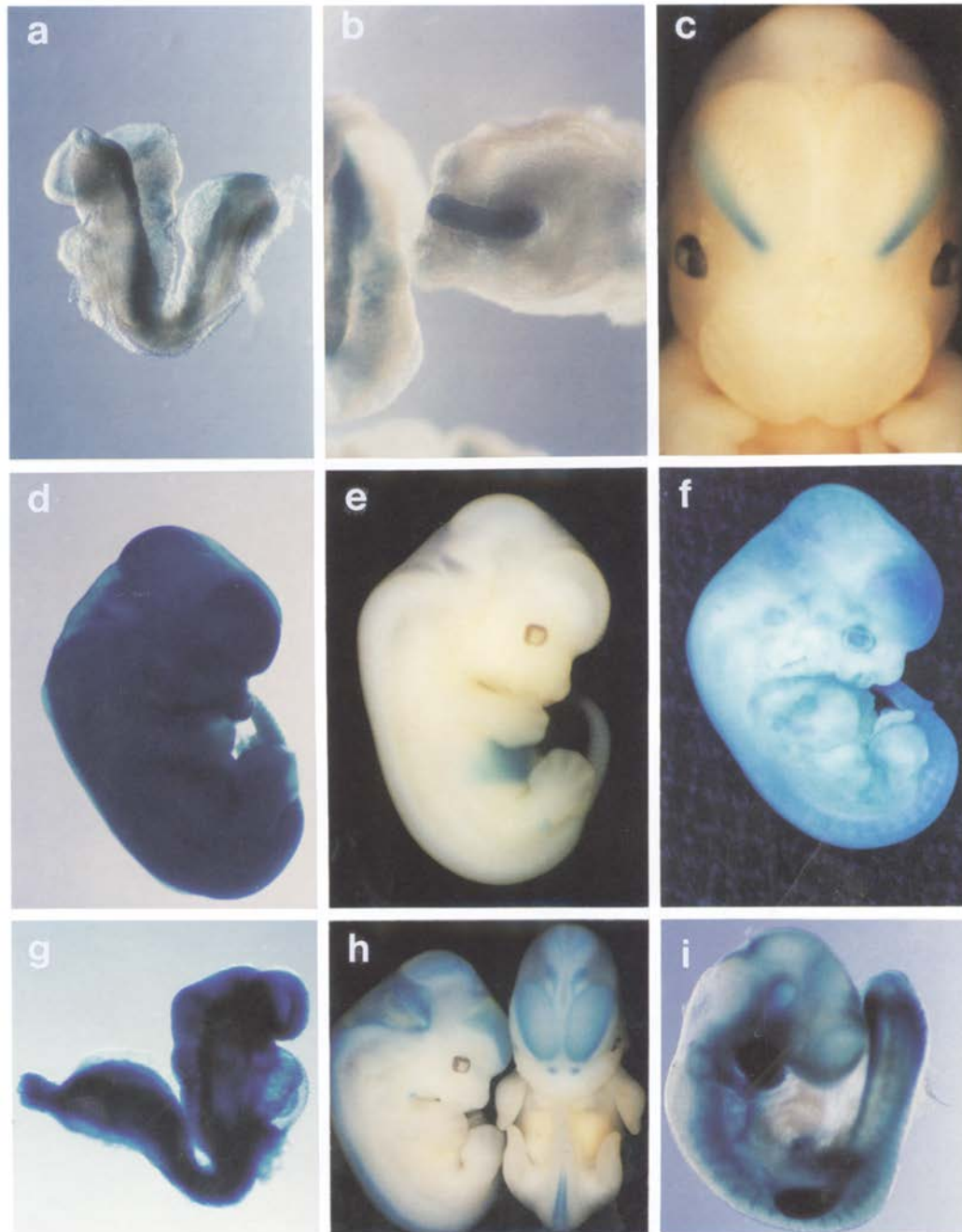
ROSA26 & its variants

- The gold standard in reporting Cre activity
- A “promoter trap” – similar to the concept of “enhancer trap” in *Drosophila*
- The ROSA β geo26 (GtROSA26) line was initially derived from pools of ES cells infected with the retroviral gene trap vector Gen-ROSA β geo at low multiplicity of infection
- β geo encodes a bifunctional lacZ/neomycin phosphotransferase gene and the ROSA26 strain is one of several strains that exhibits broad lac Z staining.



Map of the ROSA26 locus

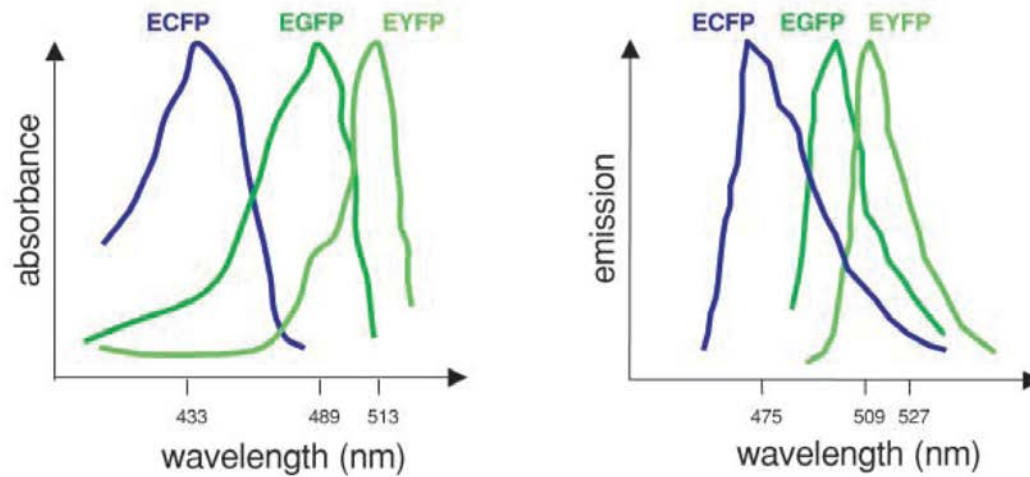




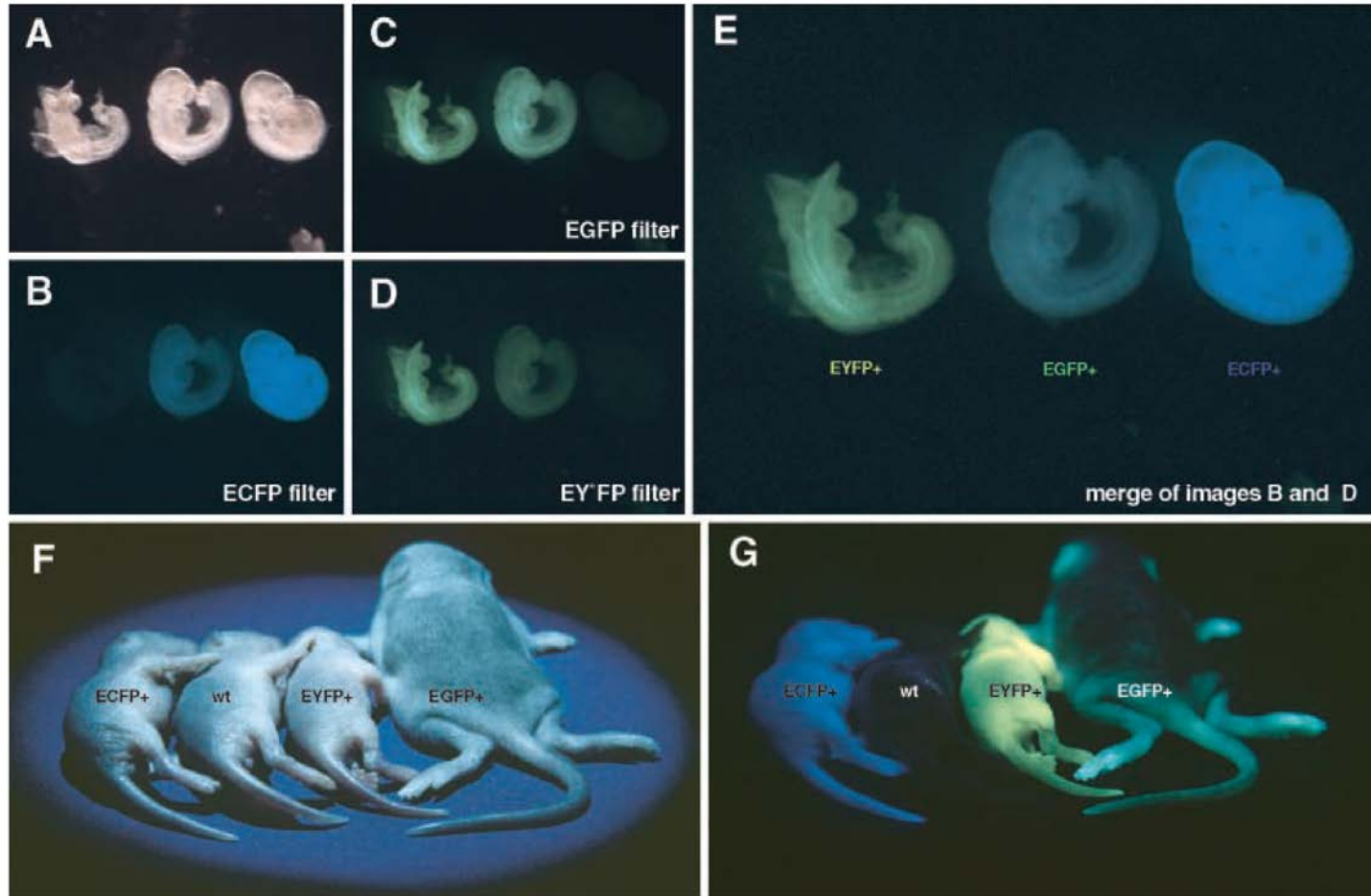
(Friedrich and Soriano, 1991)

GFP reporter and its variants

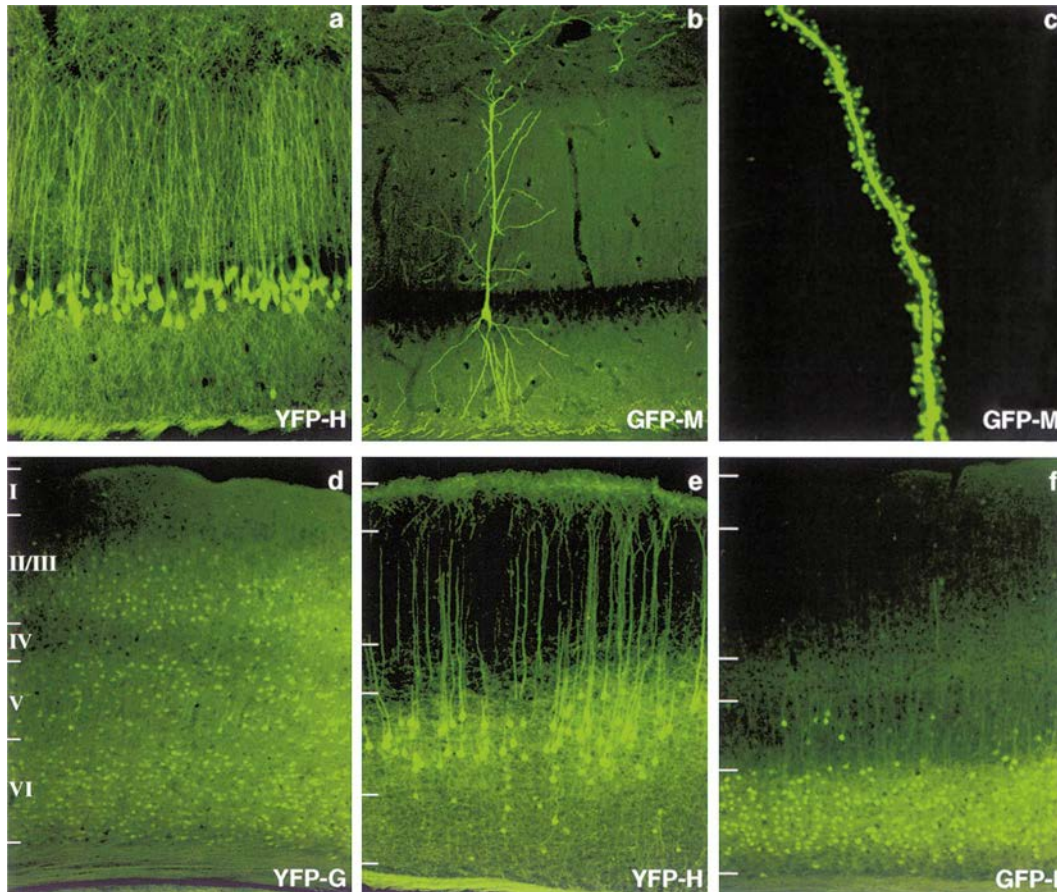
| | | |
|--------------|--|---------------------|
| wtGFP | Phe ⁶⁴ -Ser-Tyr-Gly-Val-Gln ⁶⁹ ...Ser ⁷² ...Tyr ¹⁴⁵ ...Thr ²⁰³ | Chalfie et al. 1994 |
| ECFP | Phe ⁶⁴ Leu, Ser ⁶⁵ Thr, Tyr ⁶⁶ Trp, Asn ¹⁴⁶ Ile, Met ¹⁵³ Thr, Val ¹⁶³ Ala blue shifted humanized codon usage | Heim and Tsien 1996 |
| EGFP | Phe ⁶⁴ Leu, Ser ⁶⁵ Thr red shifted humanized codon usage | Cormack et al. 1996 |
| EYFP | Ser ⁶⁵ Gly, Val ⁶⁸ Leu, Ser ⁷² Ala, Thr ²⁰³ Tyr red shifted humanized codon usage | Ormo et al. 1996 |



GFP reporter and its variants



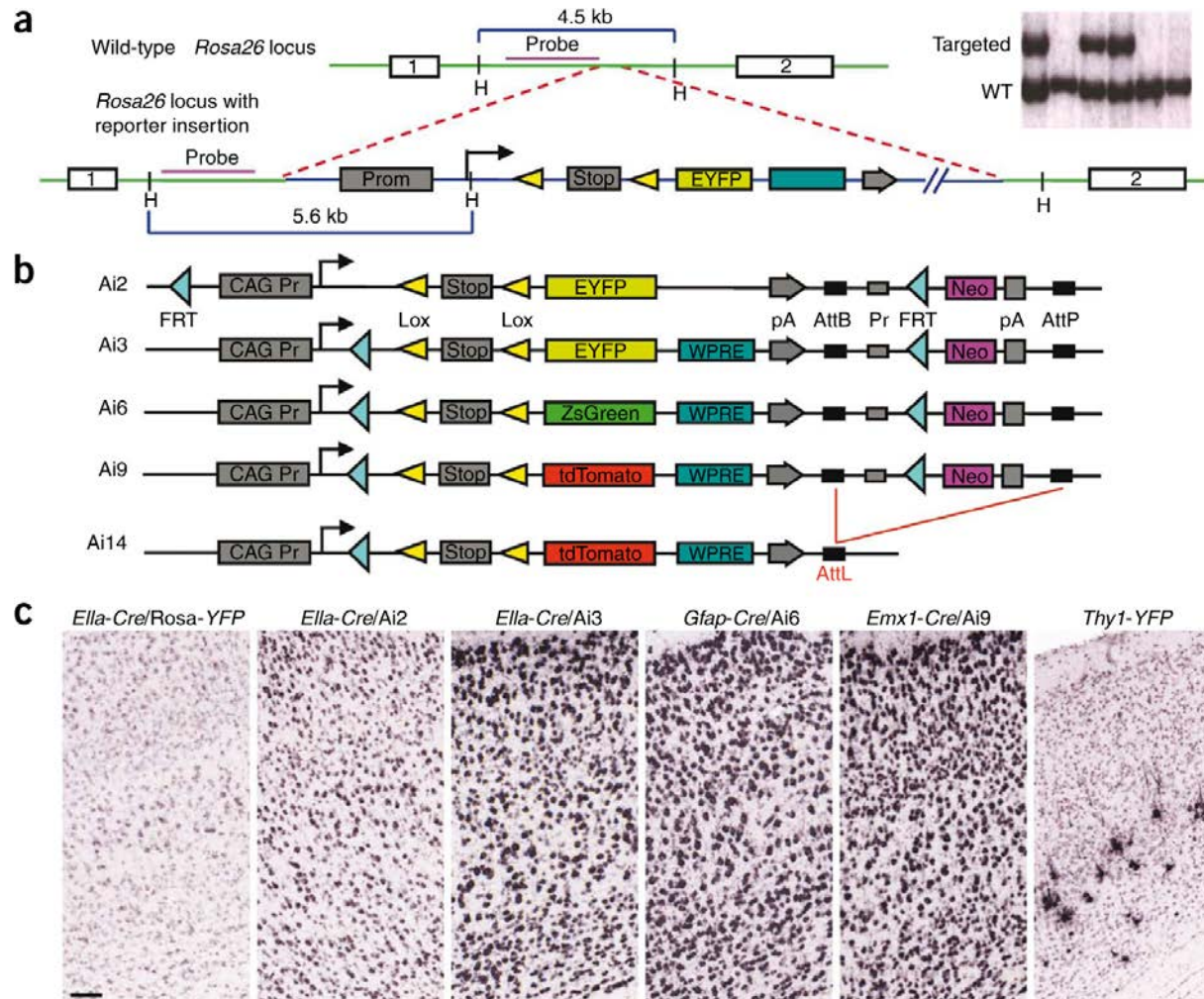
Variable expression patterns of XFP in the hippocampus and cerebral cortex



(Feng et al., Neuron 28: 41-51, 2000)

- Thy1 – a member of the Ig superfamily expressed by projection neurons in many parts of the nervous system
- Characteristics of thy1-XFP transgenic mice
 - Label axons and dendrites even though not fused to peptides designed to facilitate transport
 - Stable expression of XFP up to 9 months with no discernible effect on synaptic structure
 - Remarkable variability in patterns of XFP expression in mice generated from the same construct
 - Each line exhibit unique, heritable pattern of expression
 - Double transgenic lines can be achieved

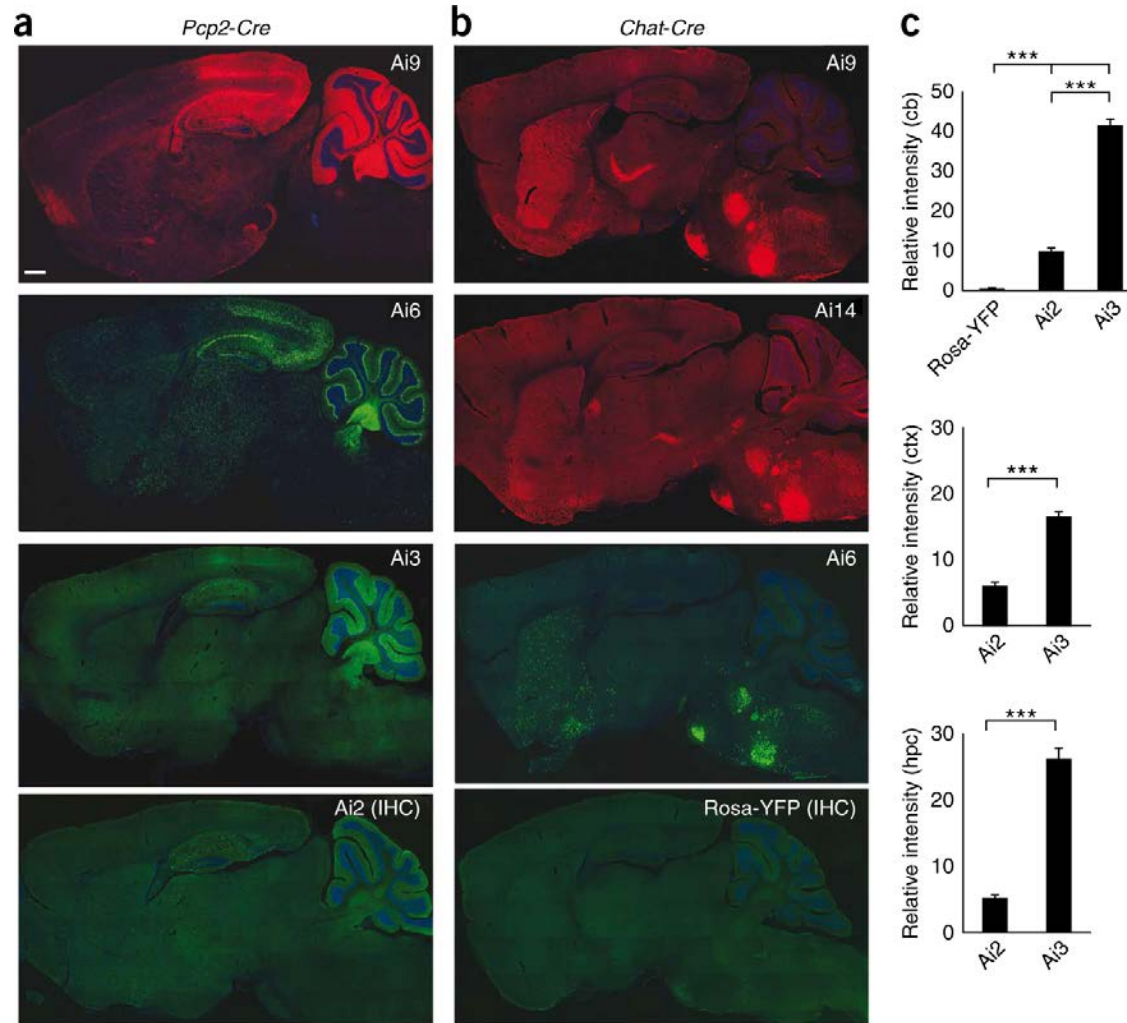
Cre reporter lines



WPRE: woodchuck hepatitis virus posttranscriptional regulatory element (to enhance RNA stability)

(Madisen et al., Nature Neurosci 13: 133-140, 2010)

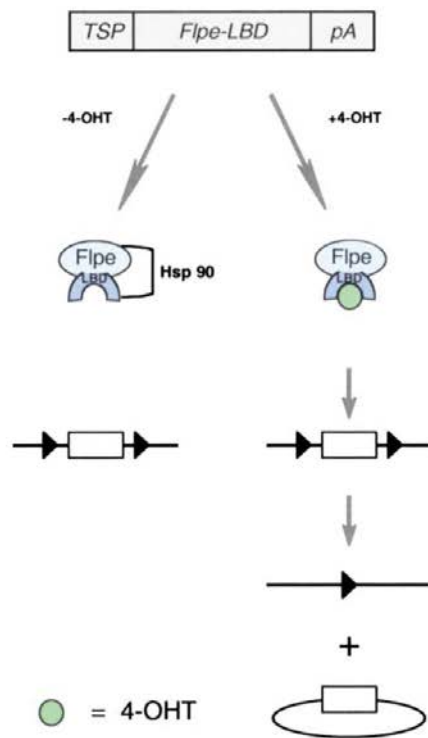
Enhanced fluorescent labeling in the new Cre reporter lines



(Madisen et al., Nature Neurosci 13: 133-140, 2010)

Mechanisms of temporal control of site-specific recombination

A Post - translational control



References

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- Komiyama, T., Johnson, W.A., Luo, L. & Jefferies, G.S. From lineage to wiring specificity: POU domain transcription factors control precise connections of *Drosophila* olfactory projection neurons. *Cell* 112: 157-167, 2003.
- Feng, G., Mellor, R.H., Bernstein, M., Keller-Peck, C., Nguyen, Q.T., Wallace, M., Nerbonne, J.M., Lichtman, J.W. & Sanes, J.R. Imaging neuronal subsets in transgenic mice expressing multiple spectral variants of GFP. *Neuron* 28: 41-51, 2000.
- Madisen, L., Zwingman, T.A., Sunkin, S.M., Oh, S.W., Zariwala, H.A., Gu, H., Ng, L.L., Palmiter, R.D., Hawrylycz, M.J., Jones, A.R., Lein, E.S. & Zeng, H. A robust and high-throughput Cre reporting and characterization system for the whole mouse brain. *Nature Neuroscience* 13: 133-140, 2010.
- Tsien, J.Z., et al. Subregion and cell type-specific gene knockout in mouse brain. *Cell* 87: 1317-26, 1996.
- Tsien, J.Z., et al., The Essential Role of Hippocampal CA1 NMDA Receptor–Dependent Synaptic Plasticity in Spatial Memory. *Cell* 87: 1328-38, 1996.

Reviews

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- Wu, J.S. & Luo, L. A protocol for mosaic analysis with a repressible cell marker (MARCM) in *Drosophila*. *Nature Protocol* 1: 2583-89, 2006.
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- Branda, C.S. & Dymecki, S.M. Talking about a revolution: the impact of site-specific recombinases on gene analyses in mice. *Developmental Cell* 6: 7-28, 2004.

2016-2017 NS201B Overview

- Signaling I: Notch signaling & lateral inhibition (Cheyette)
- Signaling II: Shh & Wnt (Cheyette)
- Neurocircuit development (Stryker)
- Signaling III: Neurotrophic factors (Huang)
- Growth cone dynamics (Weiner)
- Dendritogenesis (YN Jan)
- Synapse formation I (Ullian)
- Problem set I (2nd students)
- Synapse formation II (Ullian)
- Synapse homeostasis (Davis)
- Cortical development I (Rubenstein)
- Cortical development II (Pleasure)
- Cortical development III (Alvarez-Buylla)

NS201B 2016-2017

- Neurodevelopment & behavior (Manoli)
- In vitro models & neuropsychiatric diseases (Willsey)
- Paper discussion – Cortical development (Pleasure)
- Neurogenetics I (Sanders)
- Neurogenetics II (Ptacek)
- Myelination/Oligodendroglia/Schwann cells (Chan)
- Astrocytes (Molosky)
- Problem set II (2nd year students)
- Glia paper discussion (Molosky)
- Final Exam (1/18/17 – 1/20/17)
- ***Topics to be covered by minicourses:*** “Neurobiology of disease” (2016), “Glia biology” (2016), “Neuroinflammation” (2016), “Addiction” (2017), “Thalamocortical circuit” (2017), “Basal ganglia” (2017), etc