

**Receptor pharmacology in neuroscience practice**  
**Lecture 1: basic terms, experimental approaches and caveats**

# **Ionotropic vs. metabotropic neurotransmitter receptors**

## **What is an ionotropic receptor?**

### **Also called a ligand-gated ion channel**

Transmembrane proteins which allow ions to pass through in response to binding to a chemical messenger/ligand/neurotransmitter

E.g. GABA-A, glycine (anionic -); nAChR, ZAC, 5HT3 (Cationic +): Cys-loop receptors

AMPA, NMDA, kainate: iGluRs (cationic)

P2XRs: ATP gated channels (cationic)

Kir: PIP2 activated K<sup>+</sup> channels

**Different from voltage gated channels,  
which pass ions depending on membrane potential**

# **What is a metabotropic receptor (GPCR)?**

**Also called a G protein coupled receptor**

Transmembrane proteins which signal through second messengers to ion channels in response to binding to a chemical messenger/ligand/neurotransmitter

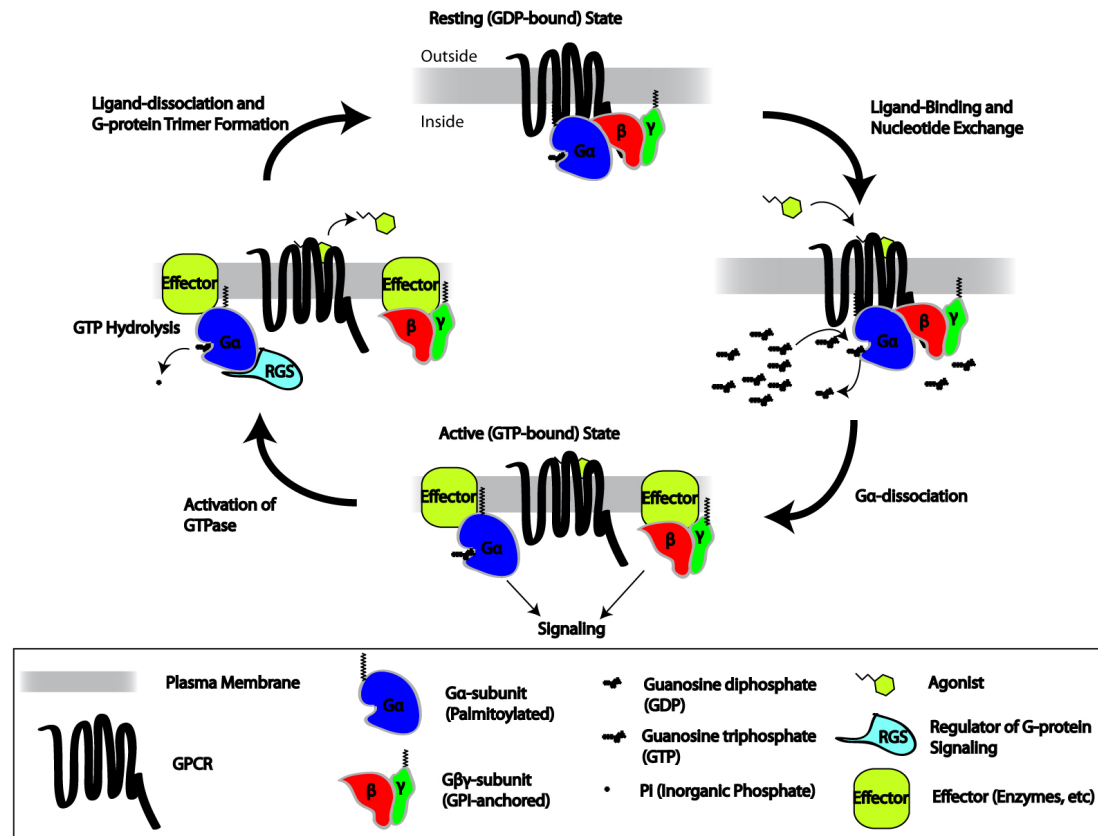
E.g. GABA-B, mGluRs, mAChR, 5HTRs, P2YRs....hmm, these look familiar

As well as norepinephrine, epinephrine, histamine, dopamine, endocannabinoids, adenosine and neuropeptides such as opioids, somatostatin, neurokinins, oxytocin, bradykinin...

**Many ligands can act on both ionotropic and metabotropic Receptors...e.g. glutamate, acetylcholine, serotonin, ATP**

# What is a metabotropic receptor (GPCR)?

It's a G protein coupled receptor...so it couples to G proteins  
G proteins are trimeric molecules...with  $\alpha\beta\gamma$  subunits.



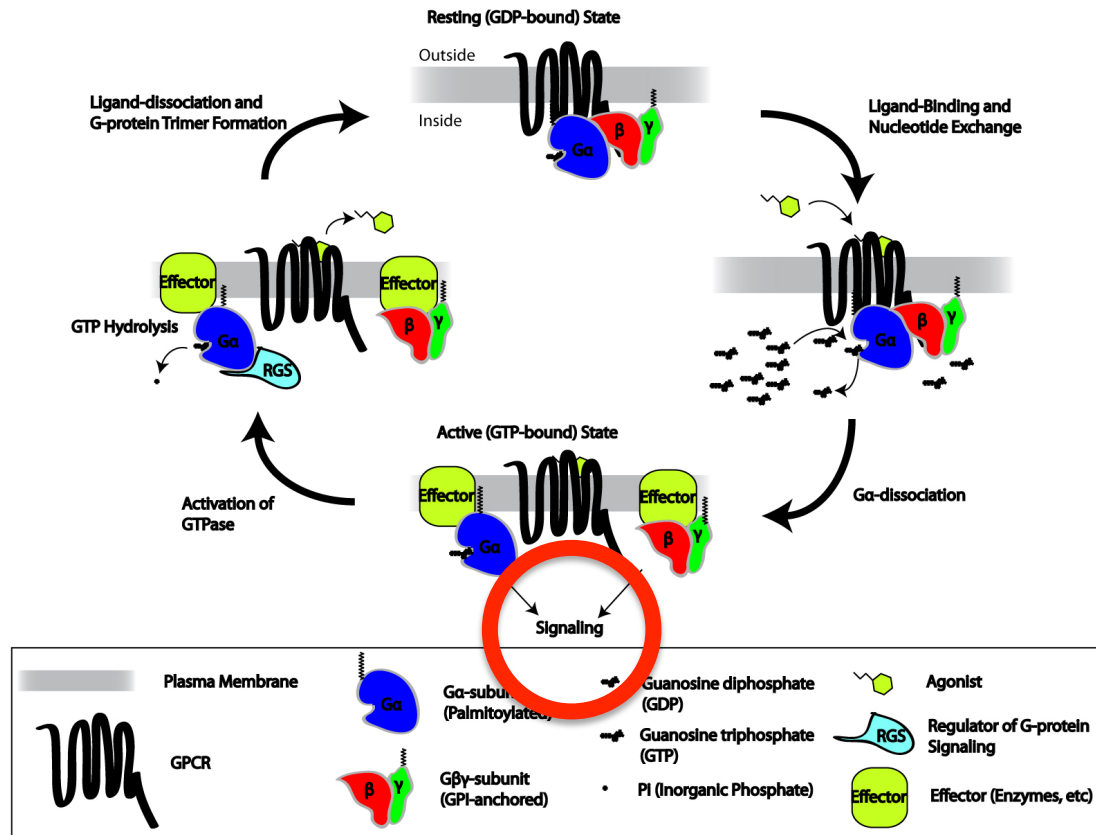
# G proteins come in several flavors defined by the $\alpha$ subunit :

G $\alpha$  i/o

G $\alpha$  s/olf

G $\alpha$  q

G $\alpha$  12/13



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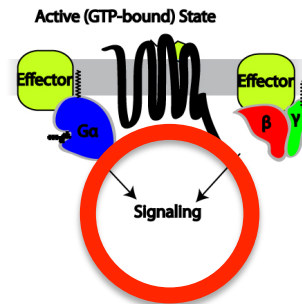
**G $\alpha$  i/o** : “inhibitory”; inhibits AC (and PKA),  $\uparrow$  K<sup>+</sup> channels,  $\downarrow$  Ca<sup>2+</sup> channels

**G $\alpha$  s/olf** : “stimulatory”; activates AC (and PKA),  $\uparrow$  cation channels

**G $\alpha$  q** :  $\uparrow$  PLC $\beta$ , converts PIP<sub>2</sub> to DAG and IP<sub>3</sub>; activating PKC and releasing Ca<sup>2+</sup> from stores

**G $\alpha$  12/13**: activate RhoGEFs, Rho, and ROCK (cytoskeletal rearrangements)

By subunits from G<sub>i/o</sub> also  $\uparrow$  GIRK K<sup>+</sup> channels and  $\downarrow$  P/Q- and N -type voltage gated Ca<sup>2+</sup> channels



There is also G protein independent signaling from GPCRs...more on that Monday....

# Pharmacology Terms You Need to Know

**LIGAND** A molecule that binds to the receptor

**AGONIST** A ligand that activates the receptor

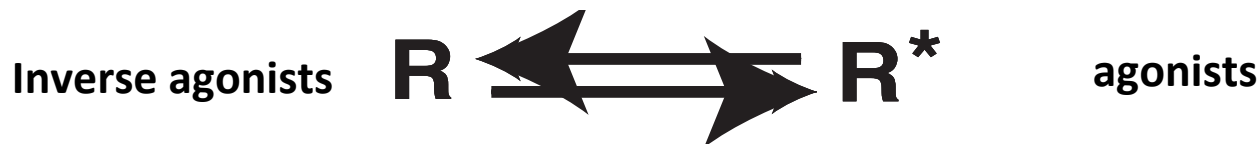
**Full agonist** An agonist that under the specified condition produces the maximal response

**Partial agonist** An agonist that under the specified condition produces a response less than that of a full agonist

**ANTAGONIST** A molecule that blocks the effects of an agonist

**Neutral antagonist** Blocks the effects of an agonist (competitive and non competitive) but doesn't alter the "basal" equilibrium of the receptor

**Inverse agonist** Blocks the constitutive activity of a receptor



The concentration dependence of the effect is a function of ligand **AFFINITY**

The magnitude of the effect is dependent on ligand **INTRINSIC EFFICACY**

Ligand **POTENCY** is a function of both of these

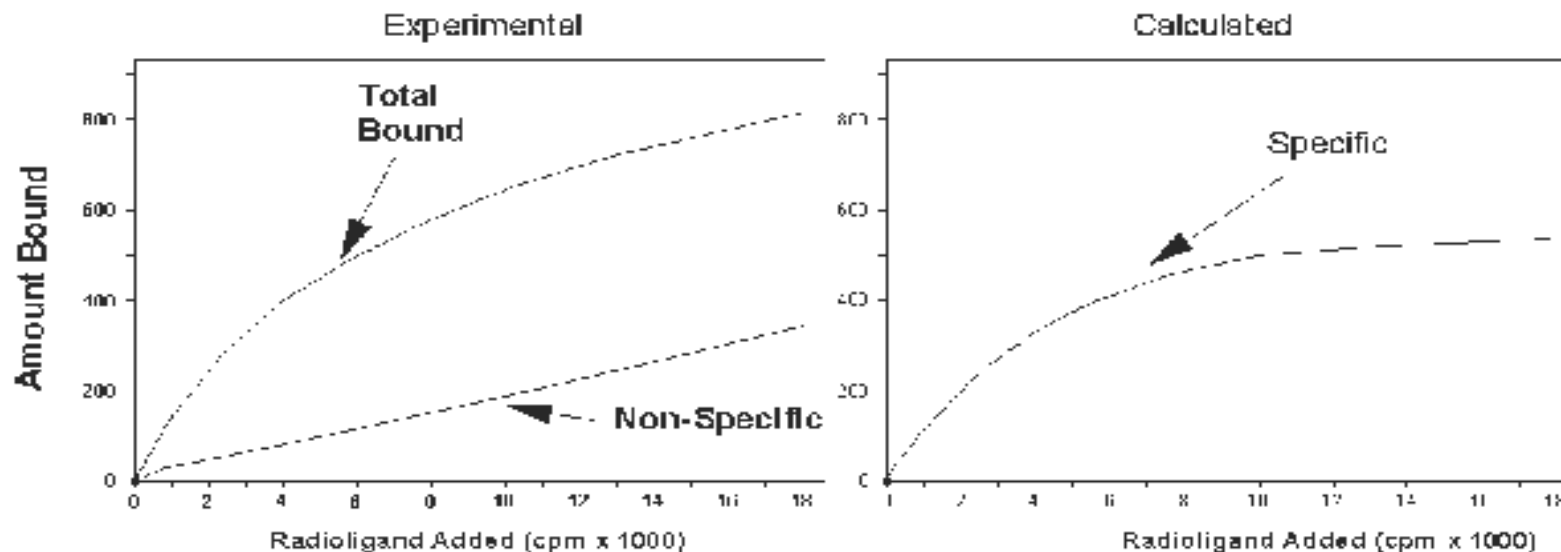
# Determining Ligand Affinity

**AFFINITY:** The ability of a ligand to bind a receptor

Expressed as a dissociation constant,  $K_d$ , in molar units  
where a lower number is higher affinity

$K_d$  is the concentration of ligand in which half the receptors are occupied.  
It is determined experimentally using “saturation binding” with a radioligand  
(or substitute).

Radioligands bind specifically to receptors and stick to other things too. Specific binding  
Is determined experimentally by subtraction of non-specific binding.



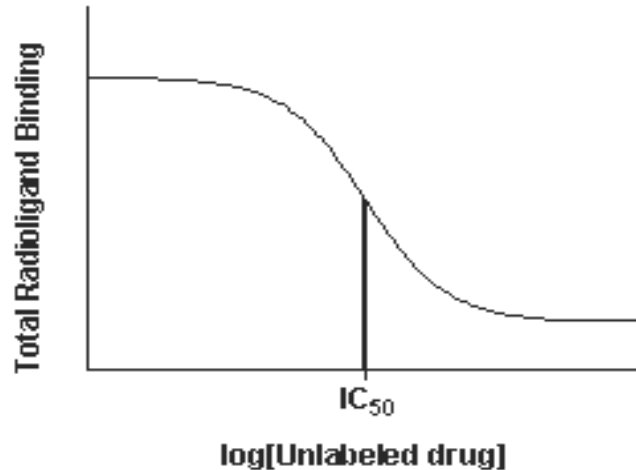
Saturation binding also gives you the total receptor number “ $B_{max}$ ”



# Determining Ligand Affinity in practice

Most reported “affinities” are determined using competition binding, not saturation binding. They give a  $K_i$  not a  $K_d$ .

Competition binding uses a fixed concentration of a radioligand (the “tracer”) and competes binding of that tracer with a range of concentrations of cold ligand



Competition Binding gives an  $IC_{50}$  (or  $EC_{50}$  value)

From this value, and the  $K_d$  of the tracer, one calculates a  $K_i$  value for the new ligand

The  $K_i$  is the concentration of competing ligand that would bind half of the receptors in the absence of any other competition.

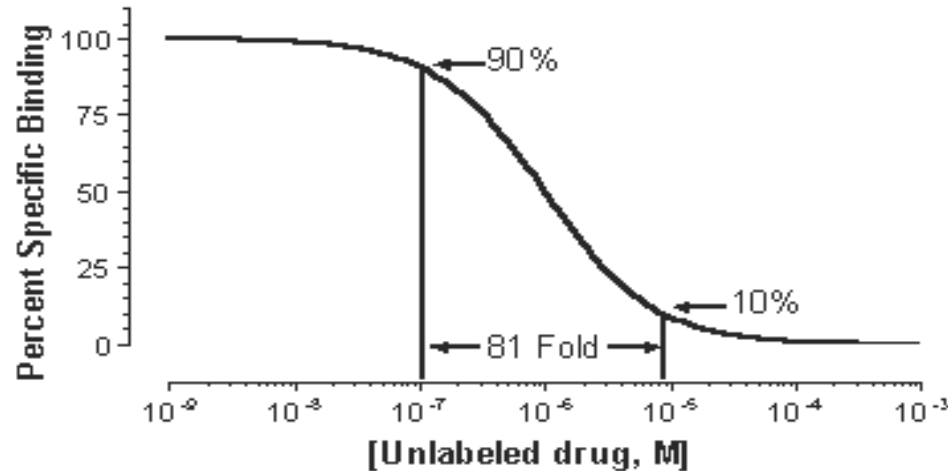
For pure competition at a single binding site  $K_i$  is :

$$K_i = \frac{IC_{50}}{1 + \frac{[ligand]}{K_d}}$$

From Cheng and Prusoff (Biochem. Pharmacol. 22: 3099-3108, 1973) there are many equations for varying situations)

# What to expect in competition binding

If labeled and unlabeled ligand compete for a single site, the competition binding curve should have a shape defined by the laws of mass action

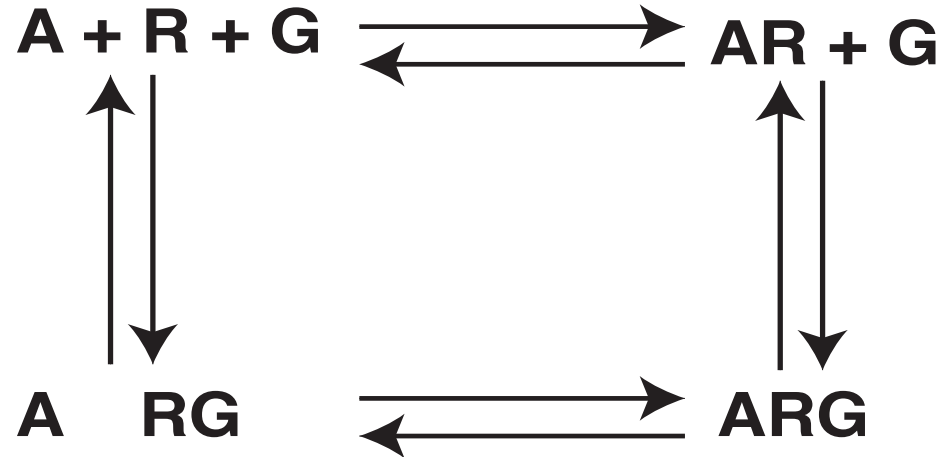


The steepness of the curve is the “Hill slope” which should be  $\sim -1$

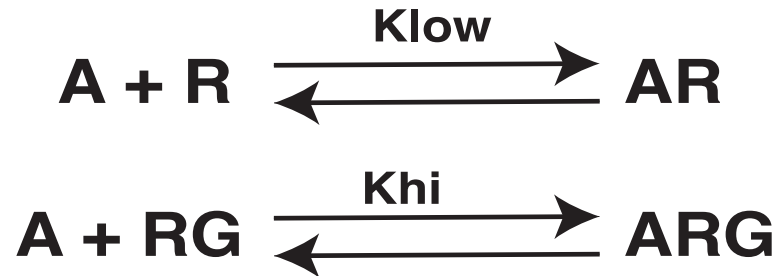
For an agonist competing an antagonist this is almost never the case  
--affinity is affected by the presence of G protein activation

In addition, If your system contains more than one binding site with different affinities the Hill slope will be shallow or biphasic (if affinities are very different)

# Agonists have differing affinities depending on G protein coupling



**An oversimplified model calculates both the low and high affinity states**  
(this is normally what is reported if high and low are calculated)



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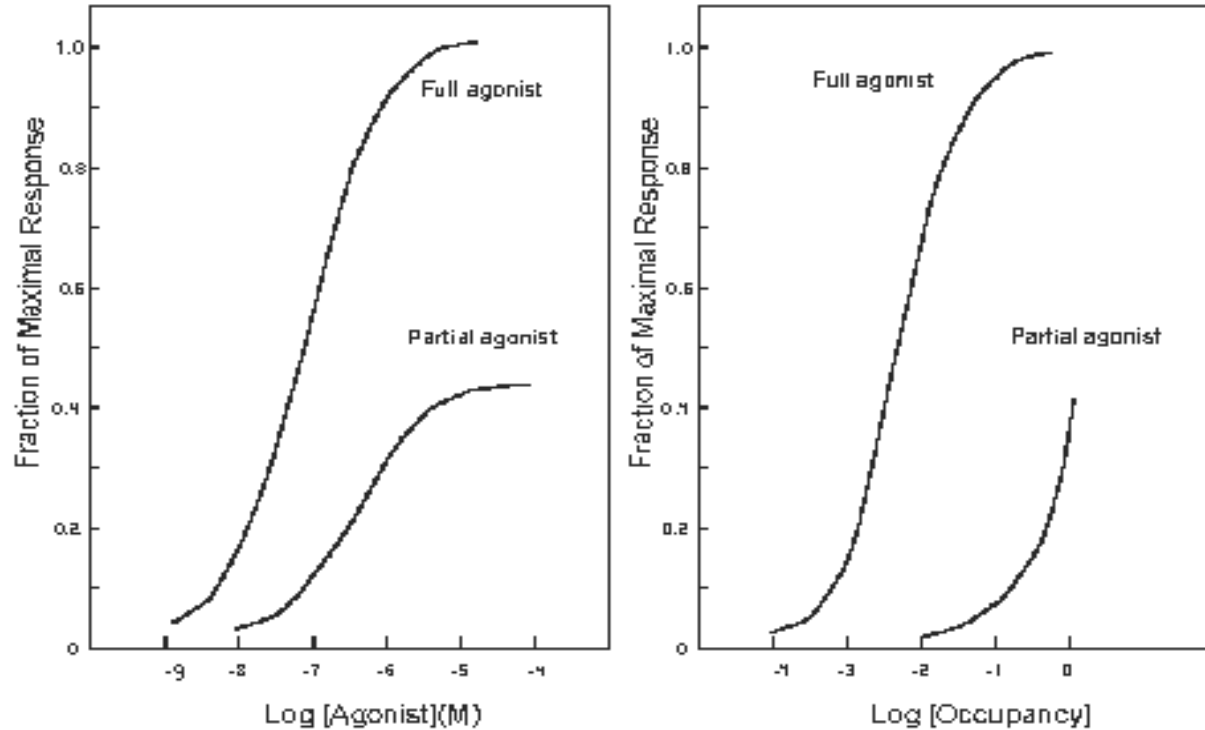
Ligand **POTENCY** is a function of both of these

# Intrinsic Efficacy and Partial Agonism

Intrinsic efficacy is the relative ability of a ligand to produce the maximal effect possible

**Caveat: Relative to what? In what system/context?**

**Relative to endogenous ligand...In a system/context you care about.**



**Intrinsic efficacy is expressed as “Emax (or % Emax relative to a standard).”**

# Intrinsic Efficacy and Partial Agonism

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**What are some things that could affect the intrinsic efficacy of your ligand?**

## **Receptor number**

Ligand X needs only 100 occupied receptors to produce maximal response, ligand Y needs 10,000. In systems with only 100 receptors available to talk to your readout Y will look like a partial agonist, but in systems with lots of “spare” receptors talking to your readout, Y will perform like a full agonist (at high enough concentration).

## **Membrane potential (or potential at which spike is fired, or inhibited)**

In cell X, the resting potential is -40 so you need only 100 receptors occupied by a full agonist to activate enough Ca<sup>2+</sup> channels to fire a spike, but I need 10,000 occupied by the partial agonist to do the same job.

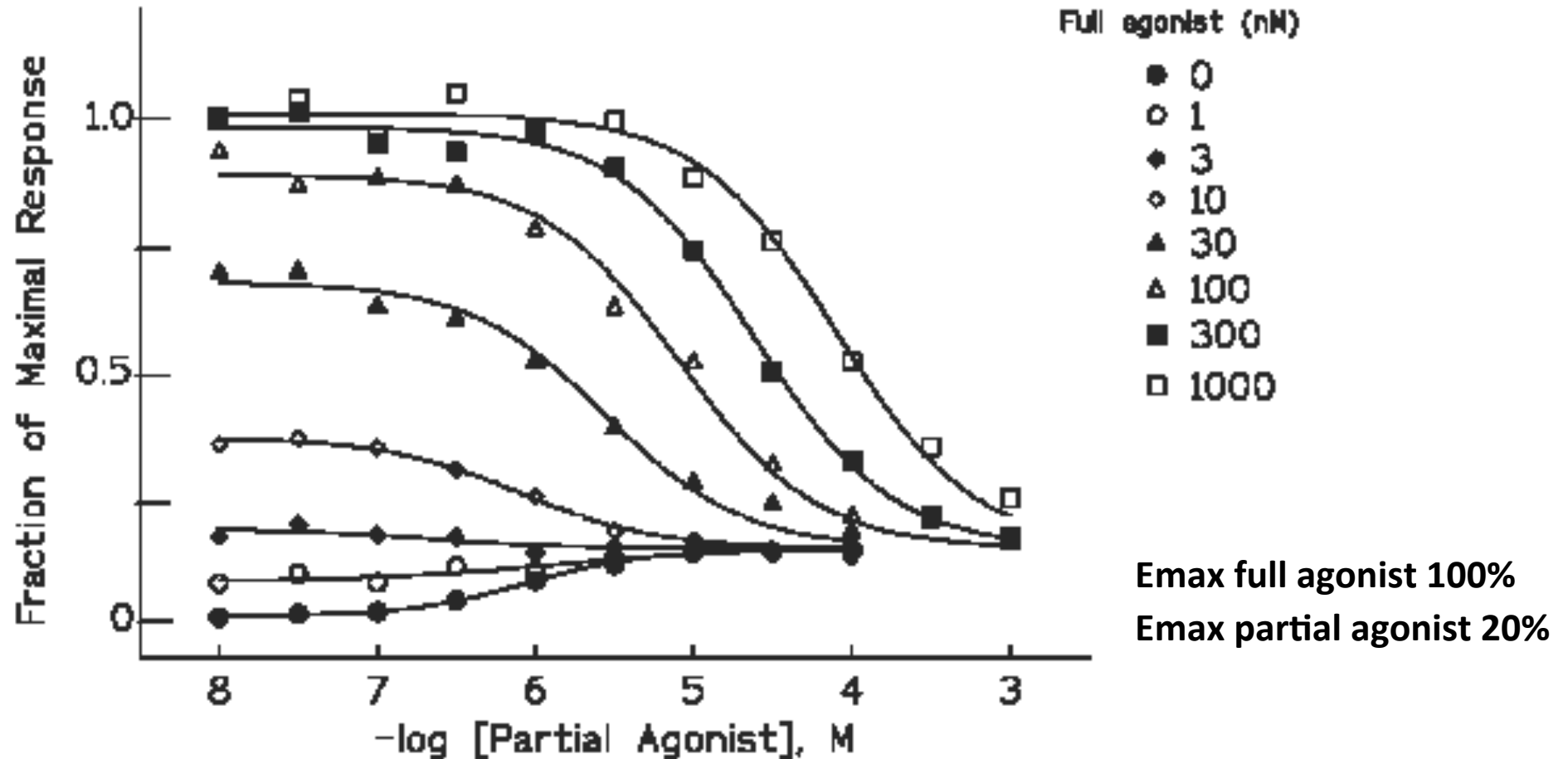
What would happen in cell Y where the resting potential is -80?

## **What other receptors/channels are engaged while ligand is around**

Especially relevant for systems that use the same ligand for ionotropic/metabotropic receptors, or in systems/preparations where you aren't controlling release of other transmitters (glutamate, GABA, glycine), or when comparing effects on pre- vs. post-synaptic responses to the same ligand.

# Intrinsic Efficacy and Partial Agonism

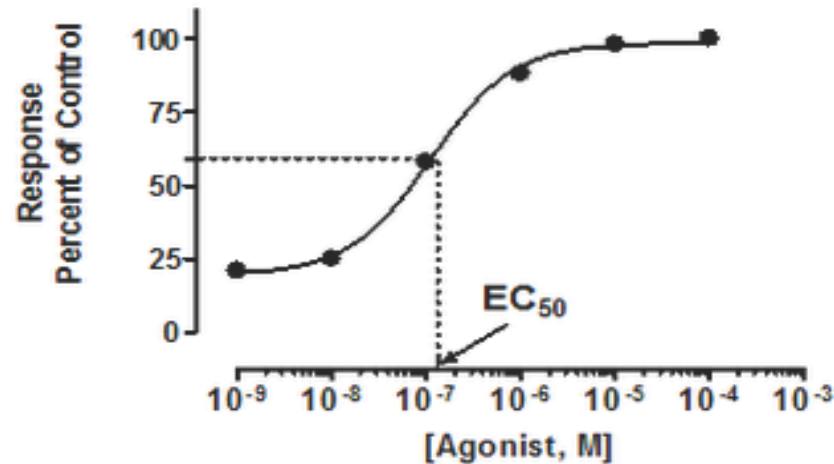
Partial agonists can work as functional antagonists for full agonists (especially if they are high affinity)



Beware: in vivo you rarely know the concentration of your endogenous ligand

# Ligand Potency and Selectivity

The EC<sub>50</sub> (for agonist) and the IC<sub>50</sub> (for antagonist) is the concentration of ligand necessary to produce 50% of the maximal effect.

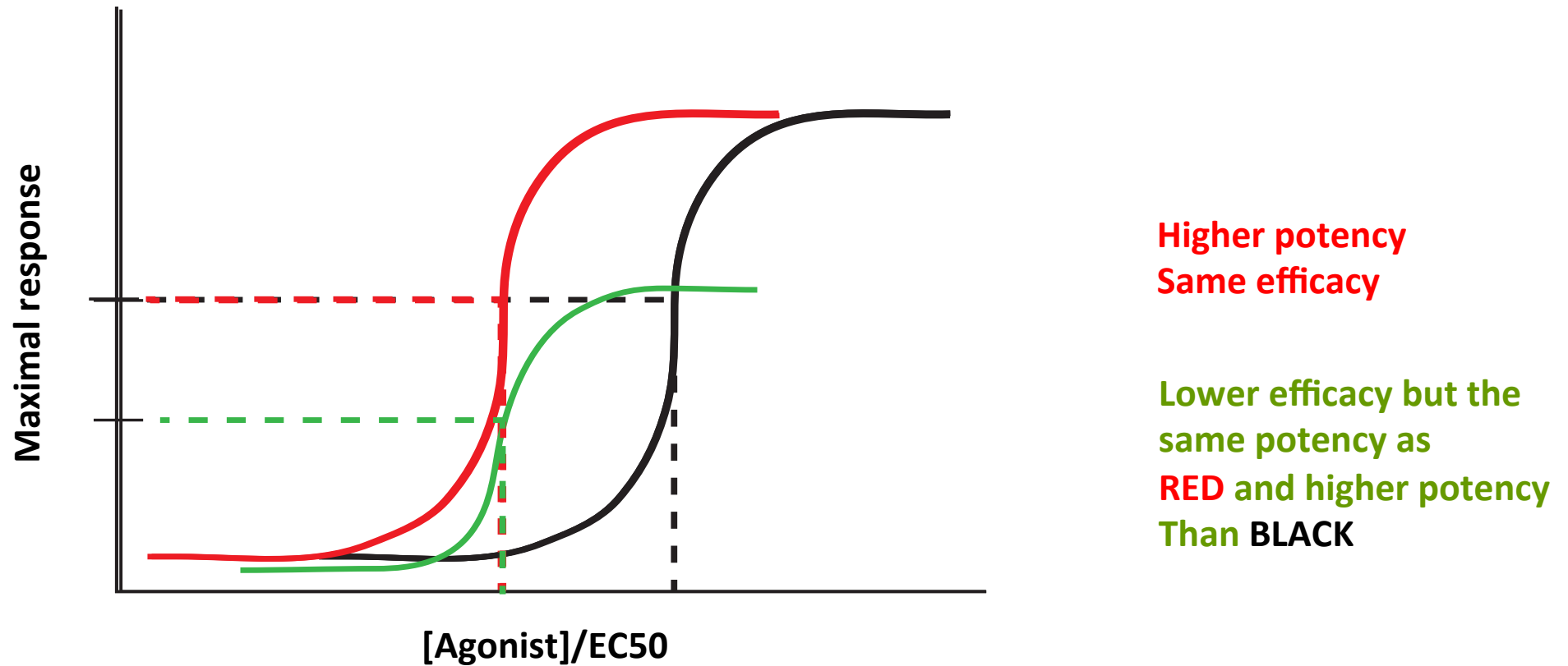


Potency is dependent on BOTH affinity and efficacy of the ligand

In most cases, potency is expressed as 50% maximal response to that ligand not to a reference ligand (for example the endogenous ligand)



# Ligand Potency and Selectivity



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**So how is “selectivity” determined?**

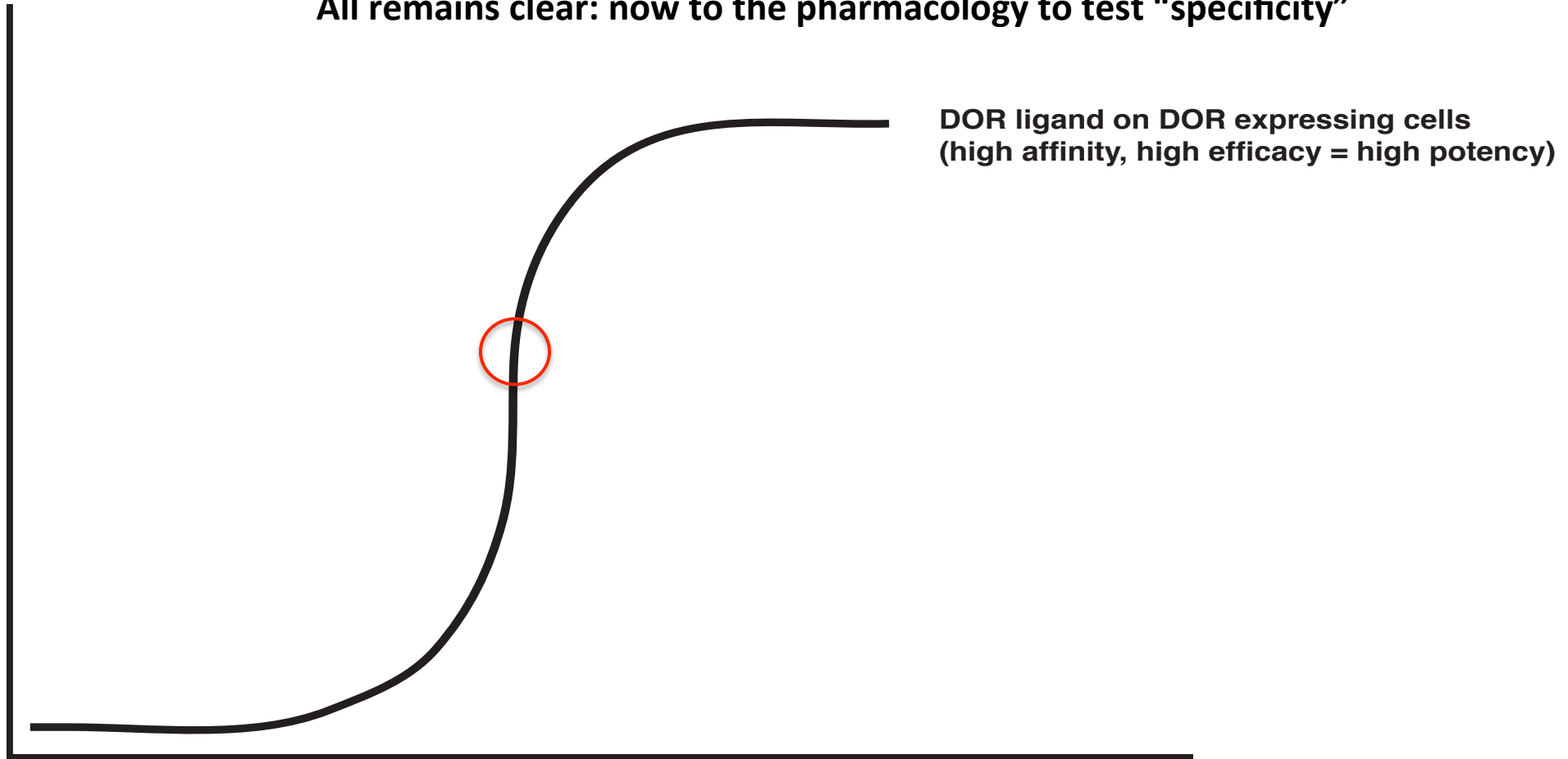
# Ligand Potency and Selectivity

**SELECTIVITY:**

**Caveats:**

# The danger of relying on “selectivity”

All remains clear: now to the pharmacology to test “specificity”



# The danger of relying on “selectivity”

## Expected effects on “DOR” expressing cells

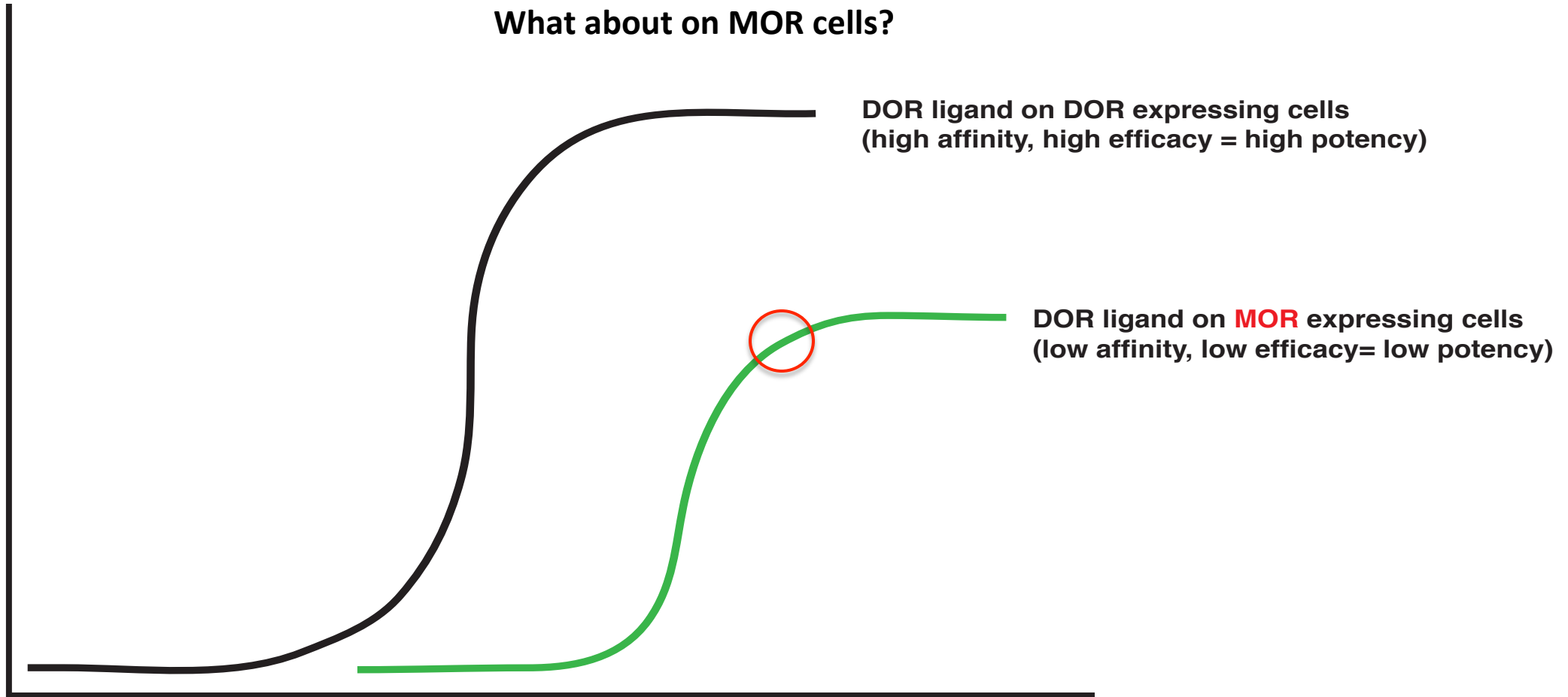
On DOR cells all is still clear



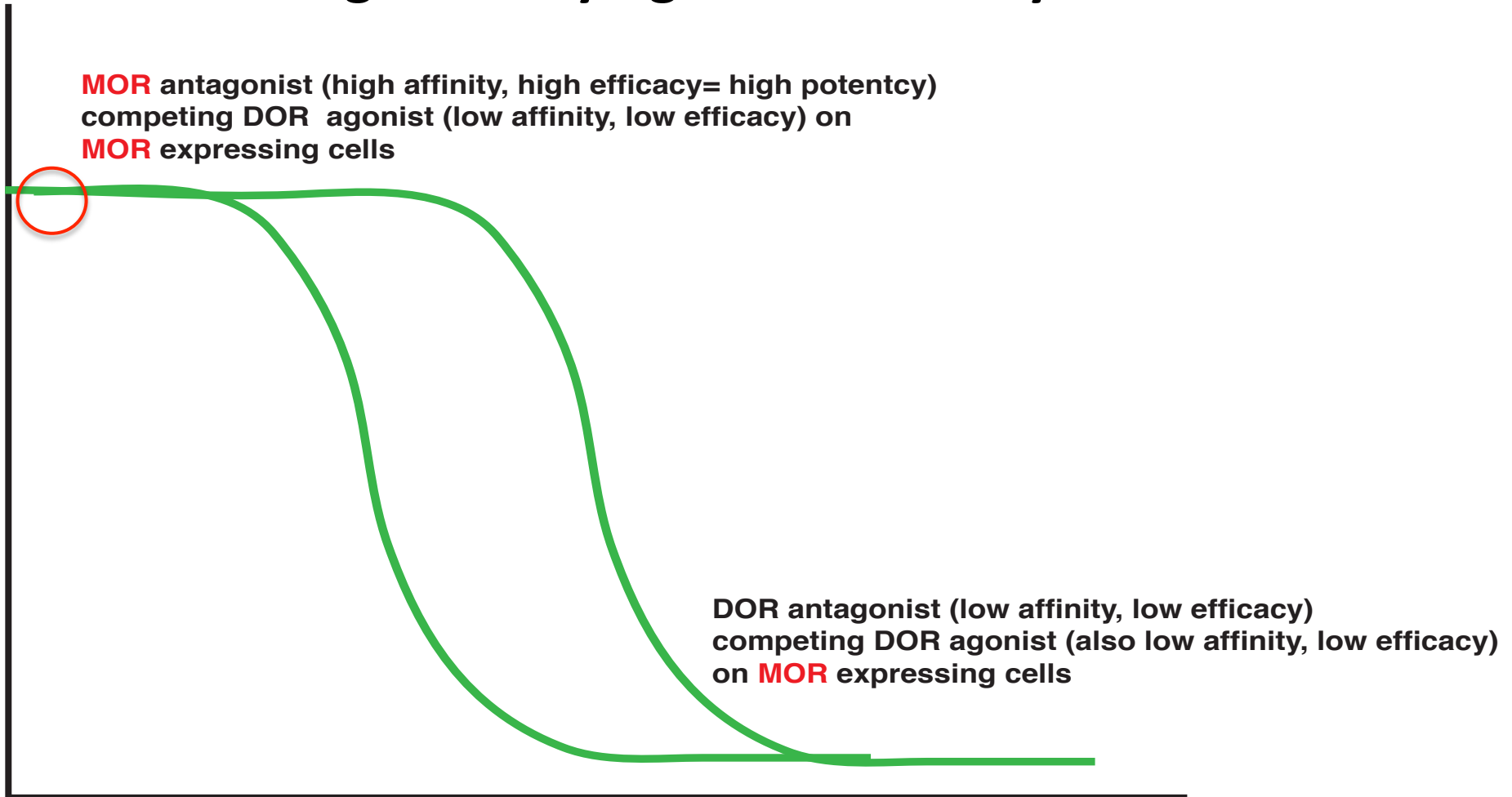
DOR antagonist (high affinity, high efficacy)  
competing DOR agonist (also high affinity, high efficacy)  
on DOR expressing cells

# The danger of relying on “selectivity”

What about on MOR cells?



# The danger of relying on “selectivity”



When you are reading the literature...

pay attention! Because now you know the benefits and limitations of a rich pharmacopeia and how to interpret others' data

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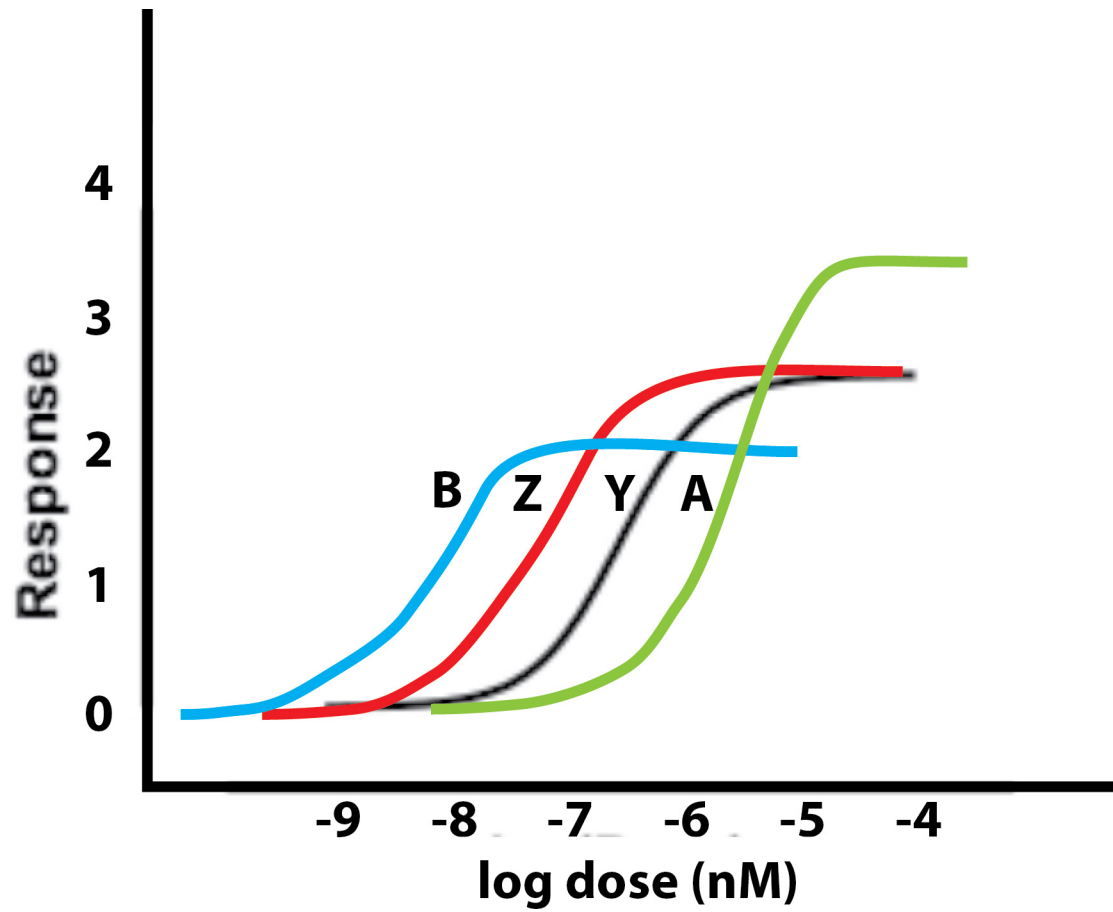
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# If you thought that was complicated.....

There is significant, mounting evidence for “Functional Selectivity”, “Biased agonism” “differential engagement” or “RAVE” (relative activity vs. endocytosis) at GPCRs...

## Functional Selectivity/Biased agonism

A single ligand can be an agonist, partial agonist, antagonist or inverse agonist depending on the effector that is being measured.

